

Original Article**Effect of Dimethyl Sulfoxide on Bond Strength of a Self-Etch Primer and an Etch and Rinse Adhesive to Surface and Deep Dentin**Farahnaz Sharafeddin ¹, Raha Salehi ², Negar Feizi ²¹Dept. of Operative Dentistry, Biomaterials Research Center, School of Dentistry, Shiraz University of Medical Sciences, Shiraz, Iran.²Postgraduate Student, Dept. of Operative Dentistry, School of Dentistry, Shiraz University of Medical Sciences, Shiraz, Iran.**KEY WORDS**

Dentin pretreatment;
 Dimethyl sulfoxide;
 Bond strength, deep dentin;
 Surface dentin;
 Etch and rinse adhesive;
 Self-etch adhesive

ABSTRACT

Statement of the Problem: Composite bond to dentin is crucial in many clinical conditions particularly in deep cavities without enamel margins due to insufficient penetration of adhesive into demineralized dentin.

Purpose: The aim of this study was to assess the shear bond strength (SBS) of a methacrylate-based and a silorane-based composite resin to surface and deep dentin after pretreatment with dimethyl sulfoxide (DMSO).

Materials and Method: Eighty extracted human premolars were randomly divided into two groups of flat occlusal dentin with different cuts as A: surface group (sections just below the dentinoenamel junction (DEJ) and B: deep group (2 mm below DEJ). Each group was randomly assigned to 4 subgroups and their samples were restored with Adper Single bond (ASB) and Filtek Z350 or Silorane system Adhesive (SA) and Filtek P90 composite resins, using a 3×3mm cylindrical plastic mold. following these steps, the subgroups were assigned as Subgroup A₁: surface dentin+ Silorane System Primer (SSP)+ Silorane System Bonding (SSB)+ P90; Subgroup A₂: surface dentin+ 37% etchant (E37%) + Adper Single Bond (ASB)+ Z350; Subgroup A₃: surface dentin+ DMSO+ SSP+ SSB+ P90; Subgroup A₄: surface dentin+ E37%+ DMSO+ ASB+ Z350; Subgroup B₁: deep dentin+ SSP+ SSB+ P90; Subgroup B₂: deep dentin+ E37%+ ASB+ Z350; Subgroup B₃: deep dentin+ DMSO+ SSP+ SSB+ P90; Subgroup B₄: dentin +E37% +DMSO +ASB +Z350. The specimens were thermocycled at 5± 2/55± 2°C for 1000 cycles and then tested for SBS.

Results: Using DMSO as dentin conditioner increased SBS of ASB to deep dentin ($p < 0.001$) and SBS of SA to surface dentin ($p = 0.003$) but had no effect on SBS of SA to deep dentin ($p = 1.00$).

Conclusion: The ability of DMSO to increase SBS of ASB to deep dentin provides a basis for improving bonding of this composite resin in deep cavities.

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Received March 2015;
 Received in revised form April 2015;
 Accepted May 2015;

Cite this article as: Sharafeddin F., Salehi R., Feizi N. Effect of Dimethyl Sulfoxide on Bond Strength of a Self-Etch Primer and an Etch and Rinse Adhesive to Surface and Deep Dentin. J Dent Shiraz Univ Med Sci., 2016 September; 17(3 Suppl): 242-249.

Introduction

In mid-1960s, composite resin restorative materials were introduced to dentistry. [1] These materials have gained high popularity among dentists and patients. They have more conservative cavity designs which basically rely on the effectiveness of current dentin bonding agents. [2] However, composite resins still have

some limitations in use due to their physical properties, [3-5] polymerization shrinkage, [6] microleakage, low wear resistance and color stability, [7-8] and lower bond strength to dentin compared with enamel. [9] These properties are related to each other somehow; polymerization shrinkage can cause composite debonding, [6] leading to gap formation, microleakage, secondary cari-

es and so on. [10]

Thus composite resin bond strength to dentin plays an important role in restoration success since the major surfaces of a preparation to be bonded are in dentin, in different depths, sometimes without any enamel margins. Although enamel and dentin are both highly hydrophilic, bonding mechanism seems to be different in each substrate because of basic differences in their organic and inorganic contents. [11] Therefore, bonding to dentin presents a much greater challenge than to enamel, [12] especially in deeper dentin due to a decrease in inter-tubular dentin and an increase in tubular diameter as well as tubular fluid. [13-14]

Efforts to improve clinical performance of methacrylate-based composite resins have led to the development of new monomers such as ring-opening silorane and new filler technology such as nanofillers. [15] Filtek P90 (P90) and Filtek Z350 (Z350) are two dental composite resins on the market that have lower polymerization shrinkage [16] and improved mechanical properties, [17] respectively when compared with other composite resins. Ring-opening polymerization technique in the silorane system, like P90, is a recent important development in dental composite resins. The term silorane comes from "siloxane" and "oxirane", which combines their two advantages, i.e. high hydrophobicity from siloxane and ring-opening monomer from oxirane, to make silorane, with a self-etch and primer adhesive system. [18] The ring-opening process seems to be insensitive to oxygen-inhibiting action unlike methacrylate-based composite resins, which hinders polymerization by converting free radicals to stable species. [19] Generally, the mechanical properties of composite resins are related to their filler load; the more filler load in the composition the higher the mechanical properties such as wear resistance, modulus of elasticity and less polymerization shrinkage. [20] Z350 is a recently introduced nanofilled composite resin with 65–75 wt% of silica and zirconia nanofillers, [21] which has been claimed to have a low shrinkage due to its high filler content. [22]

Attempts have been made to enhance composite resin bond strength to dentin. One strategy is to condition the dentin with disinfectants like ozone gas, [23] oxalate desensitizer, [24] NaOCl or EDTA [25] and CHX. [22, 26]

In total etch adhesive systems, the adhesive may not reach the entire depth of demineralized dentin, [11] thus using a penetration enhancer may improve adhesive penetration into demineralized dentin. Recently Tjäderhane *et al.* [27] evaluated the effect of dentin pretreatment with dimethyl sulfoxide (DMSO: $(\text{CH}_3)_2\text{SO}$) before application of the adhesive. This compound is one of the best known penetration enhancers for medical purposes due to its dipolar amphiphilic nature and small size. [28] The potential for tissue penetration and excellent solvent properties make DMSO an attractive solvent candidate for the dental adhesives. [27] The few studies on DMSO available in dentistry are limited to its cryoprotection ability in preservation of pulpal and periodontal ligament undifferentiated cells. [29]

To the best of our knowledge no studies have been carried out on deep and surface dentin pretreatment with dimethyl sulfoxide for the analysis of the shear bond strength. The aim of this *in vitro* study was to evaluate the effect of surface and deep dentin pretreatment with dimethyl sulfoxide on shear bond strength (SBS) of a silorane-based and a methacrylate-based composite resin.

Materials and Method

Preparation of specimens

In this experimental *in vitro* study, 80 recently extracted human maxillary first premolars, without any caries and cracks, stored in 0.2% sodium azide, were used. The teeth were cleaned and mounted 2 mm below the cemento-enamel junction in self-polymerizing acrylic resin (Acropars, Iran) to simulate tooth position in the alveolar bone. The teeth were randomly divided into two groups ($n=40$), for sections of surface dentin (group A) or deep dentin (group B). The occlusal surfaces of the teeth were sectioned using diamond discs (D&Z; Germany) under water cooling to remove the occlusal enamel and reach the central groove to expose the surface dentin just beneath the dentino-enamel junction in group A and 2 mm below the central groove to meet the deep dentin in group B as in previous studies. [30] The exposed dentin surface of all the specimens was ground with 600-grit abrasive paper to create a standardized smear layer. Then the specimens in each group were randomly assigned to 4 subgroups ($n=10$), according to the type of composite resins used (two types) and pretreatment (with or without pretreatment) (Figure 1).

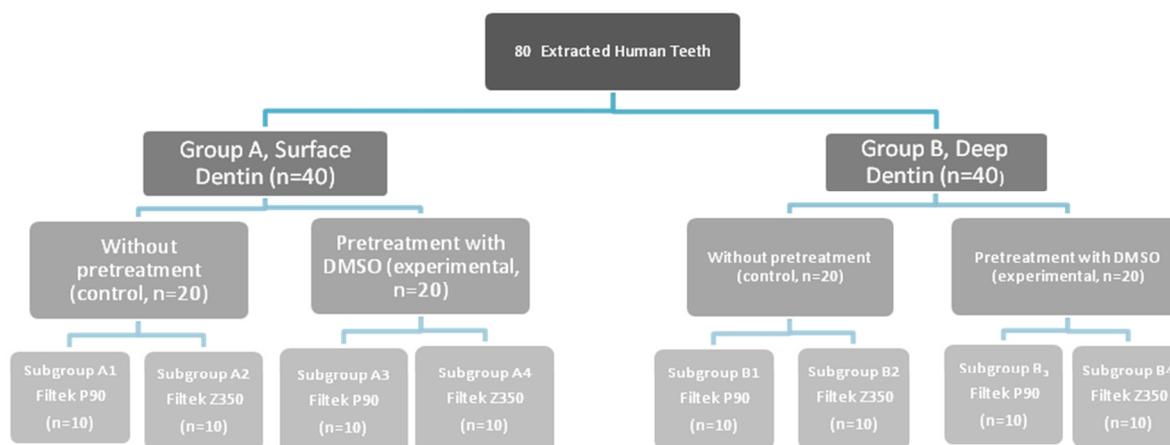


Figure 1: The study design

All the specimens were restored in accordance with the manufacturers' instructions. Composite resin build-up was done by using a 3×3-mm cylindrical translucent plastic mold, placed on the center of each specimen. A light-curing unit (Coltolux II; Coltene, USA) having a light output of 600 mW/cm² was used to cure the adhesive systems and composite resins. The light cure unit has been checked frequently after preparation of 5 teeth by using a radiometer (CM 300-1000; Halogen Lightmeter, China). The details of tested materials and their ingredients are summarized in Table 1.

Subgroup A₁: Silorane Adhesive System primer (3M ESPE, USA) was applied with a microbrush on dentin after shaking the bottle well, gently massaged for 15 s, dried with a gentle stream of air and light-cured for 10 s. Then Silorane Adhesive System bond (3M ESPE, USA) was applied, followed by a gentle stream of air and light-cured for 10 s. Filtek P90 composite resin (3M

ESPE, USA) was placed in two increments of 1.5-mm thickness and each was light-cured for 40 s.

Subgroup A₂: Acid phosphoric etchant gel (Etchant 37; DenFil Vericom Co. LTD, Korea) was applied on dentin for 15 s, rinsed with water thoroughly for 10 s and dried for 3–5 s with a gentle stream of air, leaving the surface slightly moist. Two consecutive coats of Adper Single Bond (3M ESPE, USA) were applied and light-cured for 10 s after gentle drying. Filtek Z350 (3M ESPE, USA) was placed in two increments measuring 1.5 mm in thickness and each was light-cured for 40 s.

Subgroup A₃: Dentin was pretreated with 0.004% dimethyl sulfoxide (Merck, Germany), which was obtained from diluting 3.55 ml DMSO with distilled water to have 100 ml 0.004% DMSO, for 30 s and gently dried, followed by the steps described for subgroup 1.

Subgroup A₄: Etchant gel was applied on dentin for 15 s, rinsed thoroughly for 10 s and dried with a gentle stream

Table 1: Materials, manufacturers and chemical compositions

Material	Manufacturer	Lot number	Type	Composition
Silorane Adhesive System primer	3M ESPE, USA	N469266	Two-step self-etch acid primer	Phosphorylated methacrylates, Vitrebond copolymer, Bis-GMA, HEMA, water/ ethanol solvent, silane treated silica fillers
Silorane Adhesive System bond	3M ESPE, USA	N468024	Two-step self-etch adhesive	Phosphorylated methacrylates, hydrophobic dimethacrylate, TEGDMA, silane treated silica fillers
Filtek P90	3M ESPE, USA	N496908	Silorane- based composite resin	Monomers: 3,4- epoxycyclohexyl- ethyl- cyclo- poly- methylsiloxane (5- 15 % w/w), bis-3,4- epoxycyclohexyl- ethyl- phenyl- methylsilane (5- 15 % w/w) Fillers: SiO ₂ , YF ₃ (55% v, 76% w)
Etchant 37%	DenFil Vericom Co. LTD, Korea	ET426137		H ₃ PO ₄ 37%
Adper Single Bond	3M ESPE, USA	N389084	Two-step etch-and-rinse adhesive	Bis- GMA, HEMA
Filtek Z350	3M ESPE, USA	N495372	Methacrylate-based composite resin	Monomers: Bis-GMA, UDMA, TEGDMA, PEGDMA, bis- EMA Fillers: Zirconia/ silica (63.3% v, 78.5% w)
Dimethyl sulfoxide	Merck, Germany			3.55 ml DMSO was diluted with distilled water to have 100 ml 0.004% DMSO

of air, leaving the surface slightly moist. Dentin was pretreated with 0.004% DMSO for 30 s and gently dried, followed by the bonding and restoration steps described for subgroup 2.

Subgroup B₁: The teeth were restored as previously described for subgroup A₁.

Subgroup B₂: The teeth were restored as previously described for subgroup A₂.

Subgroup B₄: The teeth were restored as previously described for subgroup A₃.

Subgroup B₃: The teeth were restored as previously described for subgroup A₄.

The teeth were stored in distilled water for 24 h at 37°C to allow for water sorption and postoperative polymerization. The teeth were then thermocycled (PC300; Vafaei, Iran) in distilled water at 5±2/55±2°C for 1000 cycles with a dwell time of 30 s.

There was a significant interaction effect between composite resin type and dentin depth ($p=0.001$), composite resin type and DMSO pretreatment ($p=0.005$) but not between dentin depth and DMSO pretreatment ($p=0.82$). We conducted a Tamhane post hoc test to compare the subgroups.

Shear bond strength analysis

Each specimen was individually subjected to a shear load applied by a wedge-shaped blade with a thickness of 1 mm in a universal testing machine (ZO20; Zwick/Roell, Germany) at a crosshead speed of 1 mm/min until failure occurred (Figure 2).

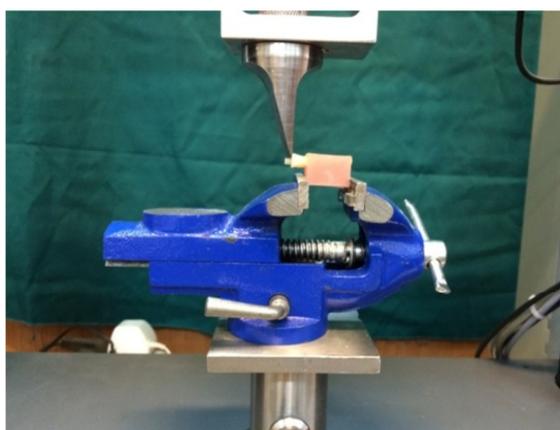


Figure 2: Shear bond strength test

The force (Newton) at failure was recorded and the shear bond strength values (MPa) were calculated from the peak load at failure divided by the bonded surface area in mm² ($A=\pi r^2, 3.14 \times 1.5 \times 1.5 = 7.065 \text{ mm}^2$).

Statistical analysis

Three-way analysis of variance (ANOVA) and Tamhane post hoc test were conducted by SPSS 15. Statistical significance was set at 0.05.

Results

The mean values and standard deviations of shear bond strengths are summarized in Table 2.

Table 2: The mean values and standard deviations of SBS

Sub-group	Definition	Mean (MPa)	Std. deviation
A ₁	Surface dentin+SA (control)	8.83 ^{AD}	1.15
A ₂	Surface dentin+ASB (control)	14.80 ^B	1.81
A ₃	Surface dentin+DMSO+SA	11.04 ^C	0.71
A ₄	Surface dentin+DMSO+ASB	16.46 ^B	2.54
B ₁	Deep dentin + SA (control)	8.21 ^{AD}	0.84
B ₂	Deep dentin + ASB (control)	9.46 ^A	0.82
B ₃	Deep dentin + DMSO + SA	7.87 ^D	1.01
B ₄	Deep dentin + DMSO + ASB	13.38 ^B	1.60

* Statistically significant difference ($p < 0.05$). The mean values followed by at least one similar letter were not significantly different.

Pretreatment with DMSO increased SBS in subgroups A₃, A₄ and B₄ in comparison with subgroups A₁, A₂ and B₂, respectively, and the increase was statistically significant in subgroups A₃ ($p=0.003$) and B₄ ($p < 0.001$) but not significant in subgroup A₄ ($p=0.963$). DMSO did not affect SBS of B₃ comparing with B₁ ($p=1.00$).

ASB exhibited significantly higher SBS values in subgroups A₂ ($p < 0.001$), A₄ ($p=0.002$) and B₄ ($p < 0.001$) than in subgroups A₁, A₃ and B₃, respectively. Subgroup B₂ had higher SBS than subgroup B₁, but the difference was not significant ($p=0.09$).

SBS was higher in all the surface subgroups compared with deep subgroups; it was significantly different in subgroup A₂ compared with subgroup B₂ ($p < 0.001$) and in subgroup A₃ compared with subgroup B₃ ($p < 0.001$) but not statistically significant in subgroup A₁ compared with subgroup B₁ ($p=0.99$) and in subgroup A₄ compared with subgroup B₄ ($p=0.14$).

Discussion

Dentin pretreatment with DMSO had a significant effect on SBS of ASB to either surface or deep dentin.

Several efforts have been made to improve composite resin bonding to dentin. One is to condition dentin surface with chemicals such as polyacrylic acid, [31] chlorhexidine digluconate [32] and antibacterial agents

[33] such as EDTA and NaOCl. Recently Tjäderhane *et al.* [27] evaluated the effect of 0.004% DMSO on nanoleakage and microtensile bond strength of Filtek Supreme XT to dentin. A methacrylate-based (Z350) and a silorane-based (P90) composite resin were used in this study with their respective adhesive systems: a total-etch system, Adper Single Bond (ASB), and a self-etch system, P90 adhesive bond. We used 0.004% DMSO to condition the surface and deep dentin and evaluate its effect on bond strength of SA and ASB. In this study DMSO increased SBS in the surface DMSO-treated SA subgroup, A₃, and deep DMSO-treated ASB subgroup, B₄, compared with respective control subgroups A₁ and B₂, consistent with a previous study [27] that reported DMSO can improve the resin–dentin bond. In surface DMSO-treated ASB subgroup, A₄, SBS was higher than its control subgroup B₂, and lower in deep DMSO-treated subgroup B₃ than its control subgroup B₁, but the differences were not statistically significant. DMSO is a chemical penetration enhancing [34] amphiphilic solvent, fully miscible in all the solvents used in adhesive dentistry. [27] This agent is able to penetrate biological surfaces [28] and compete with water molecules in inter-peptide hydrogen bonding of the collagen matrix. [35] This may explain why conditioning dentin surface with DMSO before adhesive application improved adhesive penetration into the exposed collagen matrix. The increase in SBS in the surface DMSO-treated ASB subgroup was not as much as it was expected and since this is the first time that the effect of DMSO on surface and deep dentin is studied, we recommend more studies with more specimens and different concentrations of DMSO and SEM microscopy to understand its behavior on different depths of dentin. On the other hand, the adhesive system of silorane is a self-etch one, exclusively used with this composite resin. [36-38]

The acidic hydrophilic primer with a pH of 2.7 is considered a relatively mild primer. [29] In the SA subgroups DMSO was applied before priming. It seems that the DMSO entrapped in large intra-tubular spaces of deep dentin (subgroup B₃) has a tendency to dilute the primer and weaken its already mild demineralization capacity, leading to lower SBS in deep DMSO-conditioned P90 specimens (subgroup B₃) than non-conditioned one (subgroup B₁). In contrast, comparison

of subgroup B₄ and B₂ did not show this compromising effect since DMSO was applied after acid-etching in ASB subgroups, as described in a previous study [27] and this can be the reason why SBS was higher in the ASB subgroups compared to the SA subgroups. Changes in the steps in which DMSO is applied, may lead to different results.

ASB exhibited higher shear bond strength values regardless of pretreatment or dentin depth. Different bond strength results have been reported with the use of total-etch and self-etch adhesives, with higher bond strength for either total-etch [39] or self-etch [40] adhesives. In the present study, ASB subgroups were restored using a total-etch adhesive system with 37% phosphoric acid, a strong acid (pH=0.03–0.05) [41] and the application of an adhesive, ASB. This technique leads to a complete removal of the smear layer, demineralization of the dentin surface and exposure of collagen fibers, [31] creating a demineralized zone measuring 5 to 8µm in thickness [42] but SA system's acidic hydrophilic primer with a pH of 2.7 leads to dentin decalcification limited to a few hundredths of nanometers. [38] After the primer is light-cured, a more viscous and hydrophobic adhesive resin, must be applied and light-activated independently. [43-45] The larger quantity and depth of tags obtained with total-etch systems may promote deeper micromechanical interlocking and greater SBS compared to self-etch systems like silorane. [44-46]

All the deep dentin subgroups presented lower SBS than the surface dentin subgroups, consistent with previous studies. [30] However, the difference was not significant in two pairs; first in the surface and deep SA subgroups without DMSO which might be because of lower etching potential in silorane primer that cannot demineralize the highly mineralized components of the surface dentin in comparison with deep dentin; and second in surface and deep DMSO-treated ASB subgroups due to high demineralizing potential of phosphoric acid utilized in ASB subgroups. Therefore, it seems that DMSO pretreatment is not as much effective on surface dentin as that on deep dentin when using ASB. Self-etch adhesives have a mild acid primer that partially demineralizes the dentin, despite of a total etch system that demineralizes the dentin to a higher extent and this can be the reason why we had higher SBS val-

ues in B₂ than in B₁ although the difference was not significant.

Due to insufficient data base on the mechanism by which DMSO affects different depths of dentin and dentin bond strength, further studies are required to evaluate its behavior in contact with acetone-, ethanol- or water-based dentin bonding agents and in different steps of a bonded restorative procedure. Furthermore, SEM and TEM analysis are recommended for better investigation of the depth of resin penetration with or without DMSO.

Conclusion

Within the limitations of this *in vitro* study, pretreatment of dentin with DMSO significantly improved SBS of ASB in comparison with SA. DMSO appeared to enhance SBS of ASB to either surface or deep dentin and the increase was much higher in deep dentin than in surface dentin. Therefore, when using ASB, DMSO might improve SBS in deep cavities with no enamel margins if the results of this study are confirmed by *in vivo* studies. In addition, DMSO could not affect SBS values in deep dentin restored with SA compared with surface dentin.

Acknowledgments

The authors thank Vice Chancellery of Shiraz University of Medical Sciences for supporting the research (Grant#92-01-03-6893). This article is based on the thesis by Dr. Sharafeddin. The authors also thank Dr. Vosoughi of the Dental Research Development Center of the School of Dentistry for statistical analysis, Dr. Nourisefat of the Chemistry Department of Shiraz University for kindly preparing and donating DMSO and Dr. Abdolrahimi for revising the English text of the manuscript.

Conflict of Interest

The authors of this manuscript certify no financial or other competing interest regarding this article.

References

[1] Bayne SC. Dental biomaterials: where are we and where are we going? *J Dent Educ.* 2005; 69: 571-585.
 [2] Miyazaki M, Tsujimoto A, Tsubota K, Takamizawa T, Kurokawa H, Platt JA. Important compositional charac-

teristics in the clinical use of adhesive systems. *J Oral Sci.* 2014; 56: 1-9.
 [3] Sharafeddin F, Alavi A, Talei Z. Flexural strength of glass and polyethylene fiber combined with three different composites. *J Dent (Shiraz).* 2013; 14: 13-19.
 [4] Sharafeddin F, Sharifi E. The effect of micro-wave/ laboratory light source postcuring technique and wet-aging on microhardness of composite resin. *Dent Res J (Isfahan).* 2013; 10: 370-375.
 [5] Cramer NB, Stansbury JW, Bowman CN. Recent advances and developments in composite dental restorative materials. *J Dent Res.* 2011; 90: 402-416.
 [6] Li J, Thakur P, Fok AS. Shrink-age of dental composite in simulated cavity measured with digital image correlation. *J Vis Exp.* 2014; 21(89). doi: 10.3791/51191.
 [7] Zimmerli B, Strub M, Jeger F, Stadler O, Lussi A. Composite materials: composition, properties and clinical applications. A literature review. *Schweiz Monatsschr Zahnmed.* 2010; 120: 972-986.
 [8] Sharafeddin F, Yousefi H, Modiri Sh, Tondari A, Safae Jahromi S. Microleakage of Posterior Composite Restorations with Fiber Inserts Using two Adhesives after ging. *J Dent (Shiraz).* 2013; 14: 90-95.
 [9] da Silva GR, Araújo IS, Pereira RD, Barreto Bde C, do Prado CJ, Soares CJ, Martins LR. Microtensile bond strength of methacrylate and silorane resins to enamel and dentin. *Braz Dent J.* 2014; 25: 327-331.
 [10] Nuttall CS, Vandewalle KS, Casey JA, Sabey KA. Bond strength of silorane- and methacrylate-based composites to resin-modified glass ionomers. *Gen Dent.* 2013; 61: 73-78.
 [11] Sharafeddin F, Nouri H, Koohpeima F. The Effect of Temperature on Shear Bond Strength of Clearfil SE Bond and Adper Single Bond Adhesive Systems to Dentin. *J Dent (Shiraz).* 2015; 16: 10-16.
 [12] Akbarian S, Sharafeddin F, Akbarian G. Evaluation of the influence of three different temperatures on microleakage of two self-etch and one total-etch adhesives. *J Contemp Dent Pract.* 2015; 16: 178-182.
 [13] Kinney JH, Balooch M, Marshall SJ, Marshall GW Jr, Weihs TP. Atomic force microscope measurements of the hardness and elasticity of peritubular and intertubular human dentin. *J Biomech Eng.* 1996; 118: 133-135.
 [14] Pashley DH. Clinical correlations of dentin structure and function. *J Prosthet Dent.* 1991; 66: 777-781.
 [15] Lien W, Vandewalle KS. Physical properties of a new sil-

- orane-based restorative system. *Dent Mater.* 2010; 26: 337-344.
- [16] Koliniotou-Koumpia E, Kouros P, Dionysopoulos D, Zafiriadis L. Bonding strength of silorane-based composite to Er-YAG laser prepared dentin. *Lasers Med Sci.* 2015; 30: 509-516.
- [17] Jayanthi N, Vinod V. Comparative evaluation of compressive strength and flexural strength of conventional core materials with nanohybrid composite resin core material an in vitro study. *J Indian Prosthodont Soc.* 2013; 13: 281-289.
- [18] Eick JD, Kotha SP, Chappelow CC, Kilway KV, Giese GJ, Glaros AG, et al. Properties of silorane-based dental resins and composites containing a stress-reducing monomer. *Dent Mater.* 2007; 23: 1011-1017.
- [19] Park JK, Lee GH, Kim JH, Park MG, Ko CC, Kim HI, et al. Polymerization shrinkage, flexural and compression properties of low-shrinkage dental resin composites. *Dent Mater J.* 2014; 33: 104-110.
- [20] Sharafeddin F, Bahrani S. Load Bearing Capacity of Fragmented Incisal Edges Restored with two Different Positions of Fiber Reinforced Composite Restoration. *Supplement Winter 2011*; 11: 23-28.
- [21] Leprince J, Palin WM, Mullier T, Devaux J, Vreven J, Leloup G. Investigating filler morphology and mechanical properties of new low-shrinkage resin composite types. *J Oral Rehabil.* 2010; 37: 364-376.
- [22] Hashemi Kamangar SS, Ghavam M, Mahinfar N, Pourhashemi SJ. Effect of 38% carbamide peroxide on the microleakage of silorane-based versus methacrylate-based composite restorations. *Restor Dent Endod.* 2014; 39: 172-179.
- [23] Magni E, Ferrari M, Papacchini F, Hickel R, Ilie N. Influence of ozone application on the repair strength of silorane-based and ormocer-based composites. *Am J Dent.* 2010; 23: 260-264.
- [24] Sadek FT, Pashley DH, Ferrari M, Tay FR. Tubular occlusion optimizes bonding of hydrophobic resins to dentin. *J Dent Res.* 2007; 86: 524-528.
- [25] Tjäderhane L, Nascimento FD, Breschi L, Mazzoni A, Tersariol IL, Geraldini S, et al. Strategies to prevent hydrolytic degradation of the hybrid layer-A review. *Dent Mater.* 2013; 29: 999-1011.
- [26] Breschi L, Mazzoni A, Nato F, Carrilho M, Visintini E, Tjäderhane L, et al. Chlorhexidine stabilizes the adhesive interface: a 2-year in vitro study. *Dent Mater.* 2010; 26: 320-325.
- [27] Tjäderhane L, Mehtälä P, Scaffa P, Vidal C, Pääkkönen V, Breschi L, et al. The effect of dimethyl sulfoxide (DMSO) on dentin bonding and nanoleakage of etch-and-rinse adhesives. *Dent Mater.* 2013; 29: 1055-1062.
- [28] Marren K. Dimethyl sulfoxide: an effective penetration enhancer for topical administration of NSAIDs. *Phys Sportsmed.* 2011; 39: 75-82.
- [29] Kaku M, Kamada H, Kawata T, Koseki H, Abedini S, Kojima S, et al. Cryopreservation of periodontal ligament cells with magnetic field for tooth banking. *Cryobiology.* 2010; 61: 73-78.
- [30] Villela-Rosa AC, Gonçalves M, Orsi IA, Miani PK. Shear bond strength of self-etch and total-etch bonding systems at different dentin depths. *Braz Oral Res.* 2011; 25: 109-115.
- [31] Tonial D, Ghiggi PC, Lise AA, Burnett LH Jr, Oshima HM, Spohr AM. Effect of conditioner on microtensile bond strength of self-adhesive resin cements to dentin. *Stomatologija.* 2010; 12: 73-79.
- [32] Sharafeddin F, Farhadpour H. Evaluation of Shear Bond Strength of Total- and Self-etching Adhesive Systems after Application of Chlorhexidine to Dentin Contaminated with a Hemostatic Agent. *J Dent (Shiraz).* 2015; 16: 175-181.
- [33] Mohammed Hassan A, Ali Goda A, Baroudi K. The effect of different disinfecting agents on bond strength of resin composites. *Int J Dent.* 2014; 2014: 231235.
- [34] Notman R, den Otter WK, Noro MG, Briels WJ, Anwar J. The permeability enhancing mechanism of DMSO in ceramide bilayers simulated by molecular dynamics. *Biophys J.* 2007; 93: 2056-2068.
- [35] Vishnyakov A, Lyubartsev AP, Laaksonen A. Molecular Dynamics Simulations of Dimethyl Sulfoxide and Dimethyl Sulfoxide-Water Mixture. *The Journal of Physical Chemistry A.* 2001; 105: 1702-1710.
- [36] Ivanovas S, Hickel R, Ilie N. How to repair fillings made by silorane-based composites. *Clin Oral Investig.* 2011; 15: 915-922.
- [37] Maneenut C, Sakoolnamarka R, Tyas MJ. The repair potential of resin composite materials. *Dent Mater.* 2011; 27: e20-e27.
- [38] Mine A, De Munck J, Van Ende A, Cardoso MV, Kuboki T, Yoshida Y, et al. TEM characterization of a silorane composite bonded to enamel/dentin. *Dent Mater.* 2010; 26: 524-532.

- [39] Isaac SZ, Bergamin AC, Turssi CP, Amaral FL, Basting RT, França FM. Evaluation of bond strength of silorane and methacrylate based restorative systems to dentin using different cavity models. *J Appl Oral Sci.* 2013; 21: 452-459.
- [40] McLean DE, Meyers EJ, Guillory VL, Vandewalle KS. Enamel Bond Strength of New Universal Adhesive Bonding Agents. *Oper Dent.* 2015; 40: 410-417.
- [41] Sampaio RK, Wang L, Carvalho RV, Garcia EJ, Andrade AM, Klein-Júnior CA, et al. Six-month evaluation of a resin/dentin interface created by methacrylate and silorane-based materials. *J Appl Oral Sci.* 2013; 21: 80-84.
- [42] Salvio LA, Hipólito VD, Martins AL, de Goes MF. Hybridization quality and bond strength of adhesive systems according to interaction with dentin. *Eur J Dent.* 2013; 7: 315-326.
- [43] Pucci CR, de Oliveira RS, Caneppele TM, Torres CR, Borges AB, Tay FR. Effects of surface treatment, hydration and application method on the bond strength of a silorane adhesive and resin system to dentine. *J Dent.* 2013; 41: 278-286.
- [44] Duarte S Jr, Phark JH, Varjão FM, Sadan A. Nanoleakage, ultramorphological characteristics, and microtensile bond strengths of a new low-shrinkage composite to dentin after artificial aging. *Dent Mater.* 2009; 25: 589-600.
- [45] Sauro S, Pashley DH, Mannocci F, Tay FR, Pilecki P, Sherriff M, Watson TF. Micropermeability of current self-etching and etch-and-rinse adhesives bonded to deep dentine: a comparison study using a double-staining/ confocal microscopy technique. *Eur J Oral Sci.* 2008; 116: 184-193.
- [46] Sarr M, Kane AW, Vreven J, Mine A, Van Landuyt KL, Peumans M, et al. Microtensile bond strength and interfacial characterization of 11 contemporary adhesives bonded to bur-cut dentin. *Oper Dent.* 2010; 35: 94-104.