Glypican-3 Expression in Patients with Oral Squamous Cell Carcinoma

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KEY WORDS
Glypican-3; Immunohistochemistry; Squamous cell carcinoma;

ABSTRACT
Statement of the Problem: Oral squamous cell carcinoma (OSCC) is a malignant neoplasm that affect the structures or tissues of the mouth. Early diagnosis of these tumors is important to improve the outcome of treatment. Therefore, using pathological techniques based on molecular markers may be useful for optimal diagnosis and treatment. Glypican-3 (GPC3) is involved in regulation of cell proliferation and morphogenesis and is abundant during embryogenesis and organogenesis but is limited in most of adult tissues. GPC3 overexpression has a role in carcinogenesis.

Purpose: The aim of the present study was to investigate GPC3 expression in the non-neoplastic oral epithelium and oral squamous cell carcinoma.

Materials and Method: In this cross-sectional study, 45 patients with OSCC (30 males and 15 females) with a mean age 52.3 of selected from Oral Pathology Department of Shiraz Dental School were enrolled. The control group was consisted of 15 cases of normal oral epithelium. Glypican-3 expression was assessed by using immunohistochemical methods.

Results: Non neoplastic tissues were GPC3 negative. Frequency of GPC3 positivity in tumoral tissues was recorded as 73.3% (33 cases) which was significantly higher than non-neoplastic tissues (p<0.001).

The clinicopathologic features of GPC3 expression demonstrated no association with clinicopathologic parameters except tumor size.

Conclusion: GPC3 was over expressed at protein level in oral squamous cell carcinoma, but its potential use for diagnostic and therapeutic purposes required further investigation.

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Introduction
Squamous cell carcinoma is one of the most commonly malignant tumors in the head and neck regions with a locally invasive behavior and highest mortality rate due to malignancies [1]. These cancers often revealed a clinical diagnostic challenge, especially in its early stages of growth. Attempts are being made for early diagnosis and prevention of this deadly cancer to improve its outcomes. Despite the current improvements in the treatment of this disease, OSCC is a disorder with a high mortality rate and its 5-year survival rate is still poor [2], therefore, special attention has recently been focused on the use of molecular biomarkers as reliable diagnostic tools of tumors [3].

GPC3- a cell surface protein- is a member of glypican family and belongs to a group of heparin sulphate proteoglycan bound to the cell membrane through a glycosyl-phosphatidylinositol anchor [4-5]. In mam-
mals, glypican family consists of six membranous, GPC1 to GPC6 [6]. All glypicans have a common structure that includes a glycosylphosphatidyl inositol anchor to the cell membrane, an N-terminal globular cysteine-rich domain and a C-terminal heparin sulfate glycaminoglycan attachment site [4, 7].

In humans, the gene which codes GPC3 is localized to the X chromosome (Xq26) and its product can interact with different molecules such as fibroblast growth factor2, tissue factor pathway inhibitor, Wnt5a and bone morphogenetic protein 7 [8-10]. GPC3 regulates the cell proliferation and morphogenesis and is abundant and during embryogenesis and organogenesis [11], but is limited in most of adult tissues [12-14].

GPC3 overexpression has a role in carcinogenesis, particularly in hepatocellular carcinoma [9, 15-22] but recently it was apparent that GPC3 expression is involved in different extrahepatic malignant tumors including malignant melanoma [23], pulmonary squamous cell carcinoma [13], merkel cell carcinoma [24], and chromophobe renal cell carcinoma [25].

The aim of the present study was to investigate GPC3 expression in the non-neoplastic oral epithelium and OSCC by using immunohistochemical methods and to explain the relation of it expression and clinicopathologic features.

Materials and Method
In this cross-sectional study, a total of 45 OSCC paraffin blocks were selected from Oral Pathology Department of Shiraz Dental School. The control group was consisted of 15 cases of normal oral epithelium near the lesions.

Firstly, H & E slides of available blocks were evaluated to confirm the diagnosis and adequate cellular tissue were selected for immunohistochemical staining (IHC). IHC staining was performed by using Envision Labeled Peroxides System (DAKO, Carpentaria, CA, USA). All the samples have been fixed in 10% buffered formalin and embedded in paraffin. Sections with 4μ thickness were prepared, deparaffinized in xylene, rehydrated in graded alcohol and washed with distilled water. Antigen retrieval was performed using DAKO cytomation target retrieval solution with PH = 9, for 20 minutes. Internal peroxidase activity was inhibited by 3% H2O2. Tissue sections were incubated for 30 minutes with the anti-glypican-3 antibody (Abcam, ab66596) at a 1/100 dilution.

Normal samples were stained with the same amount of antibody used for staining tumoral tissues. Gastric epithelium was used as positive control. Negative control was obtained by omission of primary antibody. Brown membranous and cytoplasmic staining for glypican-3 was considered as positive.

The slides were assessed under a light microscope (Olympus CX31; Tokyo, Japan) at 400× magnification. The percentage of the positive tumor cells out of 1000 tumoral cells in 5 different fields at high magnification (x400) was calculated and the mean percentage per slide was evaluated. A tumor was considered positive for GPC3 if more than 10% of the neoplastic cells showed strong cytoplasmic and/or membranous reactivity [26].

Chi-square test was used to compare the results between the two groups and the relation with clinicopathologic features.

Results
Gender of patients with OSCC included 15 females (33.3%) and 30 males (66.7%) with a mean age of 52.3±10.7 years. Control group consisted of 8 females (53.3%) and 7 males (46.6%) with a mean age of 53.6±10.1 years. In our study, both membranous and cytoplasmic expression of GPC3 was observed in tumoral tissues (Figure 1 & 2). Non neoplastic tissues were GPC3 negative (Figure 3). GPC3 positivity in tumoral tissues was recorded as 73.3% (33 cases) which was significantly higher than non-neoplastic tissues (p<0.001).

Statistically, the expression of GPC3 was significantly higher in samples with larger tumor size (p= 0.03).

Figure 1: Cytoplasmic and membranous expression of GPC3 in oral squamous cell carcinoma (×200)
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Figure 2: Cytoplasmic and membranous expression of GPC3 in oral squamous cell carcinoma (×400).

Figure 3: Negative expression of GPC3 in normal oral epithelium (×200).

In contrast, there was no significant association between GPC3 expression and other clinico-pathological variables such as clinical staging, grading and lymph node metastasis (p > 0.05) (Table 1).

Discussion

GPC3 is an oncofetal gene that encodes a heparin sulfate proteoglycan that is anchored to the plasma membrane [4].

GPC3 has a function as a regulator of cell proliferation and morphogenesis [11] and has a key role in regulating the balance between cell death and cell growth and is involved in cell apoptosis and cell signal transduction [19, 27]. GPC3 is widely expressed in fetal and placental cells during embryonic developmental and organogenesis [27-32] but it disappears in most adult tissue under normal condition [15].

It was shown that GPC3 can be an important cause of tumorigenesis [4-5]. Recent studies demonstrate that GPC3 is a multifunctional proteoglycan molecule, which has different role in various diseases [33].

It was reported that GPC3 expression is down-regulated in lung adenocarcinoma and clear cell renal carcinoma [34-35] but overexpression is seen in hepatocellular carcinoma, ovarian clear cell carcinoma, melanoma and neuroblastoma [14, 24, 36-37].

In the present study, GPC3 overexpression was seen in OSCC in comparison to normal tissue which demonstrates the role of GPC3 in the carcinogenesis of OSCCs. Our result was in accordance with previous studies which demonstrates positive staining of GPC3 in SCC of various sites including lung, cervical, dermal, esophagus, larynx and anal canal [13, 38-39].

Aviel-Ronen et al. [38] evaluated the expression of GPC3 in lung adenocarcinoma and showed that none of the normal lung tissues stained positively for GPC3. Similar to our findings, there was no association between GPC3 expression and clinicopathological features such as age, gender, stage and outcome. In agreement with our results, another study which demonstrated the expression of GPC3 in clear cell carcinoma of ovary showed no correlation between the expression of GPC3 and clinicopathological factors, like age, gender, stage and mortality rate, except tumor size [14].

Gonzalez et al. [33] suggested that GPC-3 can inhibit cell proliferation and has a role as a tumor-suppressor gene. Currently, and the role of GPC3 in tumorigenesis and its biological functions is poorly understood and many possible mechanisms regulated by GPC3 during tumorigenesis and tumor progression should be suggested [40]. The role of GPC3 in cell proliferation and survival may be due to its interaction with insulin-like growth factor-2 [41]. Song et al. showed that Wnt signaling pathway was changed in knockout mice [10].

GPC3 can promote tumor growth by stimulation of

Table 1: Comparative GPC3 positivity in patients with different clinicopathologic characteristics.

<table>
<thead>
<tr>
<th>Variable</th>
<th>N (%)</th>
<th>Glypican-3 negative N (%)</th>
<th>Glypican-3 positive N (%)</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>T Status</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>T1+T2</td>
<td>35(77.7)</td>
<td>12(34.3)</td>
<td>23(65.7)</td>
<td>0.03</td>
</tr>
<tr>
<td>T3+T4</td>
<td>10(22.3)</td>
<td>0(0)</td>
<td>10(100)</td>
<td></td>
</tr>
<tr>
<td>N Status</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>N0</td>
<td>17(37.7)</td>
<td>3(17.6)</td>
<td>14(82.4)</td>
<td>0.2</td>
</tr>
<tr>
<td>N1</td>
<td>28(62.3)</td>
<td>9(32.1)</td>
<td>19(67.9)</td>
<td></td>
</tr>
<tr>
<td>Stage</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>I+II</td>
<td>25(55.5)</td>
<td>9(36)</td>
<td>16(64)</td>
<td>0.1</td>
</tr>
<tr>
<td>III+IV</td>
<td>20(44.5)</td>
<td>3(15)</td>
<td>17(85)</td>
<td></td>
</tr>
<tr>
<td>Grade</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>I</td>
<td>27(60)</td>
<td>9(33.3)</td>
<td>18(66.7)</td>
<td>0.2</td>
</tr>
<tr>
<td>II+ III</td>
<td>18(40)</td>
<td>3(16.7)</td>
<td>15(83.3)</td>
<td></td>
</tr>
</tbody>
</table>
canonional Wnt signaling via making a complex with Wnt molecules [8].

GPC3 can also regulate developmental growth via interaction with the hedgehog signaling pathway [42]. It can also regulate Bax and Bcl2 protein which are involved in the apoptosis signaling pathways [43].

Stronger GPC3 expression in the hepatocellular carcinoma, increased epithelial-mesenchymal transition of tumoral cells via interaction with the extracellular signal-regulated kinase (ERK) signaling pathway [44].

Recent research has demonstrated that GPC3 is involved in the proliferation, differentiation, and adhesion of tumor cells, so it has a significant role in tumor growth and metastasis. GPC3 overexpression has been associated with increased tumor growth and metastatic ability [45-46]. There is another study which demonstrates the possible role of GPC3 in malignant transformation of salivary gland tumors. Higher expression of GPC3 in malignant salivary gland tumors in comparison with benign salivary gland tumors showed in this investigation. It may revealed the role of GPC3 in development and invasion of cancers [47].

In the current study, the expression of GPC3 was correlated only with tumor size. GPC3 was currently evaluated as a potential target for tumor specific therapy and immunotherapy [48-49]. Many studies demonstrated that GPC3 peptide vaccine trigger immune response in the patients with advanced hepatocellular carcinoma and the level of immune response was associated with overall survival [50-51].

GPC3 expression observed in OSCCs proposed that GPC3 derived vaccine might have an immunotherapeutic application in these tumors. Most of the GPC3 expression pattern in OSCC cases was mixed pattern (cytoplasmic and membranous).

The functional difference between two different patterns GPC3 expression (cytoplasmic and membranous) is unexplained, therefore, more studies are recommended to determine the significance of different localization of GPC3. GPC3 can be secreted from tumoral cells and be detected in the serum of the patients [52-53].

It was shown to be a valid tumor marker of hepatocellular carcinoma, which can be used for early detection of patients with hepatocellular carcinoma by blood screening [6, 52-53]. So further investigations are needed to evaluate the association between GPC3 expression and its serum level.

Conclusion

GPC3 was overexpressed at protein level in OSCC, there was no association between GPC3 expression and clinicopathologic parameters except tumor size. Therefore, its potential use for diagnostic and therapeutic purposes required further investigation.

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Conflict of Interests

The authors declare that they have no conflict of interests.

References


