A Literature Review

miRNAs Signature in Head and Neck Squamous Cell Carcinoma Metastasis: A Literature Review

Soussan Irani

Dental Research Center, Dept. of Oral and Maxillofacial Pathology, School of Dentistry, Hamadan University of Medical Sciences, Hamadan, Iran. and Lecturer at Griffith University, Gold Coast, Australia.

KEY WORDS

Head and Neck Cancer; Metastasis; miRNA

ABSTRACT

Statement of the Problem: Head and neck cancers include epithelial tumors arising in the oral cavity, pharynx, larynx, paranasal sinuses, and nasal cavity. Metastasis is a hallmark of cancer. MicroRNAs (miRNAs) are endogenous small noncoding RNAs involved in cell proliferation, development, differentiation and metastasis. It is believed that miRNA alterations correlate with initiation and progression of cancer cell proliferation or inhibition of tumorigenesis. Moreover, miRNAs have different roles in development, progression, and metastasis of head and neck squamous cell carcinoma (HNSCC). Altered expression of miRNAs could be novel molecular biomarkers for the definite diagnosis of cancer, metastatic site, cancer stage, and its progression.

Purpose: The purpose of this review was to provide a comprehensive literature review of the role of miRNAs in head and neck cancer metastasis.

Search strategy: A relevant English literature search in PubMed, ScienceDirect, and Google Scholar was performed. The keywords 'miRNA', 'head and neck', and 'cancer' were searched in title and abstract of publications; limited from 1990 to 2015. The inclusion criterion was the role of miRNAs in cancer metastasis. The exclusion criterion was the other functions of miRNAs in cancers. Out of 15221 articles, the full texts of 442 articles were retrieved and only 133 articles met the inclusion criteria.

Conclusion: Despite the advances in cancer treatment, the mortality rate of HNSCC is still high. The potential application of miRNAs for cancer therapy has been demonstrated in many studies; miRNAs function as either tumor suppressor or oncogene. The recognition of metastamir and their targets may lead to better understanding of HNSCC oncogenesis, and consequently, development of new therapeutic strategies which is a necessity in cancer treatment. Development of therapeutic agents based on miRNAs is a promising target.

Corresponding Author: Irani S., Dental Faculty, Shahid Fahmideh Street, Hamadan, Iran. Postal code: 65178-38677. Tel: +98-81-38381086 Fax: +98-81-38381085 Email: <u>sousanirani@gmail.com</u>, and <u>s.irani@griffith.edu.au</u>

Cite this article as: Iran S. miRNAs Signature in Head and Neck Squamous Cell Carcinoma Metastasis: A Literature Review. J Dent Shiraz Univ Med Sci., 2016 June; 17(2): 71-83.

Introduction

Received July 2015;

Accepted January 2016;

Received in Revised form November 2015;

Head and neck cancer is the sixth most common malignancy, comprising 6% of all cancers [1] and includes epithelial tumors arising in the oral cavity, pharynx, larynx, paranasal sinuses, and nasal cavity. Almost all of these malignancies are squamous cell carcinoma (SCC). [2] Head and neck squamous cell carcinomas (HNSCCs) are highly heterogeneous cancers. Oral squamous cell carcinoma (OSCC) is the most frequently occurring cancer in the head and neck area (90% of all cases). Metastasis is the main cause of death [3] and mostly arises in the tongue, floor of mouth, gingiva, and

buccal mucosa. [4]

Metastasis is a hallmark of cancer and is defined as the transfer of disease from one organ or part of an organ to another part that is not directly connected to it. Tumor cells penetrate into vascular or lymphatic channels and provide the opportunity to spread. The spread is dictated by local anatomy, and each site has its own pattern. [5] The process of metastasis is complex and involves sequential steps. First, the cells detach from the primary site. Then, they spread in the tissue, move away through the extracellular matrix, invade blood vessels, and settle in the microvasculature. Finally, the cells extravasate through the vessel wall and proliferate in the recipient tissue. [6] Cervical lymph node metastasis is the strongest determinant of patient prognosis in HNSCCs [7] which decreases the survival rate by about 50%. [8-9] HNSCC is not usually detected in the early stages of the disease due to the absence of clinical symptoms. [10]

MicroRNAs (miRNAs) are endogenous small non-coding RNAs; they can regulate gene expression in the post–transcriptional stage by interacting with the 3' untranslated region (3' UTR) of the target mRNA. [11] Several miRNAs regulate cancers; miRNAs play crucial roles in proliferation, differentiation, apoptosis, survival, motility, invasion and metastasis, and morphogenesis. [12] It has been shown miRNAs can be used as novel molecular biomarkers for cancer diagnosis. [13] For example, circulating miRNAs are associated with distinct metastatic site; therefore, they are powerful tools to evaluate the disease stage and progression. [14]

The purpose of this review was to provide a comprehensive literature review of the role of miRNAs in head and neck cancer metastasis.

Searching Strategy

A relevant English literature search in PubMed, ScienceDirect, and Google Scholar was performed. The keywords 'miRNA' 'head and neck' and 'cancer' were searched in title and abstract of publications; limited to 1990 to 2015. The inclusion criterion was the role of miRNAs in cancer metastasis. The exclusion criterion was the other functions of miRNAs in cancers. A total of 15221 articles were found. Among them, the full text of 442 articles were retrieved and recited. Only 133 articles met the specific inclusion criteria for this review.

miRNAs biology

In human, miRNAs have a stem-loop structure which are capped at the 5'-end and have a 3'-poly (A). [15] Functional miRNA is produced through a two-step transcript maturation process. The first step occurs in the nucleous, which is facilitated by an endogenous RNase III called Drosha, and the double-stranded RNA binding domain protein DGCR8. [16] After cleavage of both strands of the pri-miRNA transcript by Drosha, a stemloop precursor molecule (pre-miRNA) is produced. [17] Pre-miRNA molecules are then transported to the cytoplasm. [18] Finally, both strands of the pre-miRNA are cut at the base of stem-loop by the RNA III enzyme Dicer and dsRBD protein TRBP. The final product is a 22-nucleotide long molecule. [19]

miRNAs regulate the target genes

Literature shows miRNA and its target mRNA have interaction in the 5'-end of the miRNA. For target sequence recognition, the sequence complementarity between nucleotides 2-8 is essential. [20] The 3'-UTR region of the target mRNA called "seed region" in the 5'-end of the miRNA determines the mechanism by which the miRNA regulates the target. [21] Downregulation of target genes happens at the protein level but the regulation occurs at the translational level; therefore, the mRNA level remains unaffected. [22] The mechanism by which miRNAs regulate mRNA depends on the degree of miRNA complementarity with the mRNA molecule. Identification of miRNA targets is difficult because only 6-8 bases align perfectly with the target mRNA's 3'-UTR region. [23] The predicted targets for the differentially expressed miRNAs are significantly enriched for protein-coding tumor suppressors and oncogenes. A number of the predicted targets, along with the tumor suppressors and oncogenes have been confirmed experimentally. For instance, elevated expression level of miR-155 in HNSCC may explain its possible role in oral carcinogenesis [24] by targeting tumor suppressors such as APC. [25]

The role of miRNAs in regulating EMT/MET

Invasion is the initial step in the metastatic process. The mode of tumor invasion is the most significant prognostic factor for the presence of lymph node metastasis. Cases with grades 1-3 mode of invasion show a low frequency of metastasis; while, those cases with grade 4 mode of invasion show a high frequency of metastasis.

[26] Epithelial-mesenchymal transition (EMT) is characterized by down-regulation of E-cadherin a cell adhesion molecule, and acquisition of mesenchymal markers such as N-cadherin, vimentin and fibronectin to gain cell motility and invasiveness. [27] Both EMT and Mesenchymal-epithelial transition (MET) are essential for cancer progress, as EMT of primary tumor cells is a necessity for motility and invasiveness. In addition, MET is important for the final stage of metastasis when the extravasated cancer cells revert to epithelial cells and proliferate as a secondary tumor in the metastatic site. [28] Some transcription factors are the inducers of EMT and tumor metastasis including, Snail, Slug, Twist, ZEB1, and ZEB2. [29] Several miRNAs inhibit EMT such as miR-573, [30] and miR-33b. [31] Overexpression of miR-7 also suppresses migration and invasion and partially reverses EMT via targeting IGF1R in gastric cancer. Additionally, overexpression of miR-7 reduces metastasis. [32] However, some miRNAs such as miR-214, [33] and miR-544a promote EMT. [34]

The role of miRNAs in angiogenesis in cancers

Angiogenesis has an essential role in tumor progression and metastasis. Vascular endothelial growth factor (VEGF) is one of the most important factors in angiogenesis and lymphangiogenesis. [35-36] VEGF-A expression level is conversely related to the miR-126 expression level in cancers such as lung cancer. [37] The miR-17-92 cluster is positively associated with angiogenesis in some cancers; however, individual components of the miR-17-92 cluster have opposite effects on angiogenesis. [38-39]

The role of miRNAs in cell proliferation and apoptosis

MicroRNA alterations correlate with the initiation and progression of cancer cell proliferation or inhibition of tumorigenesis. For example, miR-155 is up-regulated in OSCC and its dysregulation is associated with the low level of CDC73, a tumor suppressor gene; therefore, it promotes cancer cell proliferation. [40] A previous study on OSCC cells indicated that transfecting with miR-125b or miR-100 significantly decreased cell proliferation; however, co-transfection had a greater effect on proliferation than individual transfection. [41] miR-128 also acts as a tumor suppressor and inhibits the growth of HNSCC cells. [42] Down regulation of miR-29 in OSCC [43] is a positive regulator of p53. [44] miR-15 and miR-16 are the regulators of Bcl-2, an antiapoptotic factor. Therefore, endogenous levels of miR-15 and miR-16 correlate inversely with the Bcl-2 protein level. [45] In an *in vitro* study, miR-375 induced apoptosis in HNSCC by targeting TNF-α. [46]

The role of miRNAs in cancer development and metastasis

Previous studies have indicated that dysregulation of miRNAs plays a crucial role in the progression of oral precancerous lesion from dysplasia to OSCC. For instance, miR-31 negatively controls oral leukoplakia progression through the regulation of fibroblast growth factor 3 (FGF3). On the contrary, miR-21, miR-181b, and miR-345 are up-regulated in oral dysplasia and associated with lesion severity. [47] Overexpression or subexpression of miRNAs in cancer has the ability to activate or block the target mRNAs. In a metastatic carcinoma, the miRNA expression profile has been shown to be different from a non-metastatic tumor. [48] Therefore, interference with the expression level of miRNAs has an effect on cancer prognosis. This finding provides a therapeutic use for miRNAs. [49]

Regarding the role of miRNAs in cancer, they are divided into two main groups of tumor suppressor and oncogenes (oncomirs). Altered miRNA expression depends on its role as a tumor suppressor or oncogene. [50] Furthermore, tumor suppressor miRNAs have been shown to be down-regulated in cancers; whereas, oncogene miRNAs are up-regulated. [51] Additionally, metastamir, a specialized family of miRNAs, has proand anti-metastatic effects. [52]

Tumor suppressor miRNAs involved in HNSCC metastasis

The literature review shows miR-1 is down-regulated in hypopharyngeal SCC. It has been shown miR-1 can suppress metastasis in HNSCC by targeting TAGLN2, a gene which mediates cell migration and invasion. [53] Let-7d, a member of the let-7 family of miRNAs acts as a tumor suppressor, most likely via targeting RAS. [54] A reduction of let-7d expression in OSCC tissues has been reported. Let-7d was negatively correlated to Twist, Snail, and EMT transcription markers in OSCC cell lines. In addition, let-7d was significantly decreased in regional metastatic lymph nodes of OSCC patients. [55] Combined low levels of miR-205, and let-7d expression are associated with distant metastasis. [56] Moreover, decreased let-7d expression in HNSCC is associated with poor prognosis. Both miR-17 and miR-20a are down-regulated in advanced migratory OSCC,

and their expression levels are negatively controlled by advanced TNM stage and lymph node metastasis. ITGβ8 is a direct target of both miR-17 and miR-20a. Thus, they can be used as prognostic biomarkers for OSCC. [57] The miR-29 family is significantly downregulated in HNSCC tissues and cell lines suggesting their contribution to metastasis in HNSCC through laminin γ2 (LAMC2) and α6 integrin (ITGA6). [58] A previous study demonstrated that the transforming growth factor beta (TGF- β) signaling which contributes to EMT process, [59] inhibits the expression of miR-29 family, and promotes the expression of extracellular matrix (ECM) components. [60] miR-34b plays an important role in EMT by targeting Snail. [61] In HNSCC, upregulation of miR-34 b/c has been reported. [62-63] but the target gene has not yet been identified. [63] In addition, ectopic transfection of miR-99 family (miR-100) in HNSCC cell lines decreases cell migration, suggesting the role of miR-99 family as a tumor suppressor by targeting IGF1R and mTOR signaling pathways. [64] miR-125b has also been shown to have a role in tumor suppression by inhibiting cancer cell proliferation, migration, and invasion in skin SCC by down-regulation of matrix metalloproteinase -13 (MMP13). [65] miR-125b also significantly decreases the proliferation of OSCC cells. [41] miR-126 is a negative regulator of VEGF-A and induces nodal metastasis in OSCC patients. Therefore, downregulation of miR-126 induces angiogenesis and lymphangiogenesis by targeting VEGF-A, and basic fibroblast growth factor (bFGF). [66] miR-133a functions as a tumor suppressor in HNSCC and is significantly down-regulated in HNSCC tissues. MSN, an actin binding protein, involved in cell motility, is a novel target of miR-133a. [67] miR-138, a multi-functional molecule regulator, is frequently downregulated in OSCC, which enhances cancer cell proliferation and invasion by targeting GNA12 mRNA, a proto-oncogene. [68] miR-138 is a tumor suppressor in HNSCC cells by suppressing invasion and promoting apoptosis. [69] In addition, miR-138 regulates cell migration and invasion in HNSCC cells by targeting RhoC and ROCK2. [70] The reduction of miR-138 in highly metastatic OSCCs is associated with elevated expression of RhoC and ROCK2 which enhances the activity of the Rho GTPase signaling cascade to increase metastasis. [71] miR-153 prevents EMT, returns the mesenchymal phenotype (MET phenomenon) of cells to epithelial-like cells, and decreases cellular invasive ability. Low expression level of miR-153 is related to OSCC metastasis by targeting SNAI1 and ZEB2. [72] The miR-200 family can inhibit EMT and tumor cell migration by targeting ZEB1/ZEB2. [73] Decreased expression level of miR-200c has been shown in the regional lymph nodes of HNSCC patients, and is associated with increased expression of BMI1 in primary tumors. [74] miR-222 acts as a tumor suppressor in OSCC and inhibits tumor invasion and metastasis by indirectly regulating MMP1 expression via targeting SOD2 mRNA (superoxide dismutase 2). [75] In HNSCC, miR-363 inhibits metastasis by down-regulation of podoplanin (PDPN), a proto-oncogene. [76] A low miR-375 expression level is associated with poor prognosis and distant metastasis in patients with HNSCC. [77] In addition, low expression level of miR-375 in the late stage

Dysregulated miRNA (Reference)	Expression status	Proposed target gene	
miR-1 [53]	Underexpression	TAGLN2	
let-7d [55]	Underexpression	Twist, Snail	
miR-17 and miR-20a [57]	Underexpression	ITGβ8	
miR-29 [58-59]	Underexpression	LAMC2, ITGA6, TGF-β	
miR-34 [63]	Overexpression	Unknown	
miR-99 family (miR-100 and miR-99a) [64]	Underexpression	IGFIR, mTOR	
miR-125b [41]	Underexpression	KLF13, CXCL1 and FOXA1	
miR-126 [66]	Underexpression	VEGF-A, bFGF	
miR-133a [67]	Underexpression	MSN	
miR-138 [70-71]	Underexpression	GNA12, RhoC and ROCK2	
miR-153 [72]	Underexpression	SNAI1 and ZEB2	
miR-200 [73]	Underexpression	ZEB1,ZEB2	
miR-200c [74]	Underexpression	BMI1/ZEB1	
miR-222 [75]	Underexpression	MMP-1	
miR-363 [77]	Underexpression	Podoplanin(PDPN)	
miR-375 [1]	Underexpression	Unknown	

Table 1: Summary of Suppressor miRNAs and proposed target genes

of cancer is correlated with increased invasive capabilities and metastasis of OSCC cells. [78] Table 1 summarizes the role of tumor suppressor miRNAs in HNSCC metastasis.

OncomiRs involved in HNSCC metastasis

It is reported that miR-10b is down-regulated in HNSCC cell lines and induces cell proliferation. [79] miR-10b significantly increases oral cancer cell migration and invasion. [80] Recently, miR-21 has been studied more frequently as a prognostic marker of HNSCC. It is stated that miR-21 down-regulates programmed cell death 4 (PDCD4) in OSCC. [81] miR-21 up-regulation is correlated with advanced stages of OSCC and lymph node metastasis. [82] Therefore, a positive relationship exists between high expression of miR-21 and worsened prognosis in HNSCC. In addition, increased expression level of miR-21 in tumor stroma is significantly related to worsened outcome. [83] miR-31 is up-regulated in HNSCC and activates hypoxia-inducible factor-1a (HIF-1 α) in normoxia and enhances the oncogenesis via factor inhibiting HIF-1(FIH). In fact, miR-31 functions as an oncogene only when oxygen supply is sufficient. It is believed that miR-31 also promotes some events of early stages of OSCC such as proliferation and tumorigenicity. [84] In addition, miR-31 is significantly expressed in early stage of OSSC tissue samples and its expression is associated with the lack of lymph node metastasis. [78] Increased expression level of miR-93 is associated with HNSCC lymph node metastasis. [85] In other cancer types, miR-93 promotes tumor invasion and metastasis through regulating EMT, angiogenesis, and disrupting adhesion between cancer cells and extracellular matrix (ECM). [86-87] However, the target gene of miR-93 in HNSCC is not elucidated. High expression of miR-134 is associated with nodal metastasis and mortality and miR-134 functions via targeting WW domain-containing oxidoreductase (WWOX) gene, a tumor suppressor. [88] In addition, up-regulation of miR-155 in OSCC is correlated with the histologic grade and can be used as a potential prognostic biomarker. [89] miR-155-5p induces metastasis and is associated with poor prognosis; miR-155-5p inhibitor down-regulates signal transducer and activator of transcription 3 (STAT3) by activating suppressor of cytokine signaling 1 (SOCS1). [90] miR-181b is highly expressed in OSCC patients with lymph node metastasis.

75

[91] According to a previous study, miR-181 was upregulated, during progression of oral leukoplakia to dysplasia and finally to invasive carcinoma. [47] In HNSCC, miR-211 promotes tumor invasion and metastasis by targeting transforming growth factor β type II receptor (TGFBRII). There is an inverse correlation between miR-211 and TGFBRII expression during HNSCC metastasis. [92] A previous study indicated that miR-211 was significantly expressed in OSCC tissue samples with N2 nodal metastasis and vascular invasion compared with non-cancerous tissues. [93] miR-223 plays crucial role in tumourigenesis and its function is cancer type-specific. Reduced level of miR-223 has been observed in hematopoietic malignancies; however, elevated expression of miR-223 is indicated in several types of solid tumors of head and neck area. [64] Table 2 summarizes the role of oncomiRs in HNSCC metastasis.

Table 2: Summary of oncogene miRNAs and the propose	ed
target genes	

Dysregulated miRNA (Reference)	Expression status	Proposed Target gene
miR-10b [80]	Underexpression	Unknown
miR-21 [81]	Overexpression	PDCD4
miR-31 [84]	Overexpression	FIH
miR-93 [85]	Overexpression	Unknown
miR-134 [88]	Overexpression	WWOX
miR -155 -5p [90]	Overexpression	STAT3
miR-181b [91]	Overexpression	Unknown
miR-211 [92]	Overexpression	TGFβRII
miR-223 [63]	Overexpression	Unknown

The role of miRNAs in diagnosis and prognosis of cancer

It has been shown that miRNAs can be a non-invasive diagnostic tool in detecting HNSCC and determining prognosis as they can be detected in saliva. For example, miR-125a and miR-200a are under-expressed in saliva of OSCC patients compared to healthy controls. [94] Besides, a previous study showed that the miRNA expression profile differs in different sites of the head and neck area and reflects the developmental origin of tissue; moreover, miRNAs can distinguish the cancers from benign tumors, non-neoplastic lesions and normal tissue. For instance, the expression ratio of miR-221 to miR-375 can be used to distinguish tumor from normal tissue with high specificity and sensitivity [95] and altered expression level of 22 miRNAs can accurately distinguish chronic pancreatitis from pancreatic cancer. [96] Besides, the different expression profile of miR-

NAs in esophageal squamous cell carcinoma is a diagnostic and prognostic factor as well as a tool for distinguishing cancer from normal tissue. [97] Moreover, miRNAs help to predict the primary site of metastatic disease. [11] In head and neck area, the primary site is unknown in up to 10% of metastatic squamous cell carcinomas. By knowing the origin of the disease, specific therapeutic regimens such as reducing the radiation field can be employed, which consequently minimizes the morbidity. [98] Interestingly, the miRNA expression level in primary tumors and their corresponding metastatic sites remains consistent in different patients; however, it differs in the same tumor in different locations. All together, the miRNA expression profile can be used as a diagnostic tool to distinguish the cancers from benign tumors and other benign conditions and also from normal tissues, as well as to predict the origin of unknown primary tumors in the head and neck area. [98] Decreased expression levels of miR-205 and let-7d are significantly associated with higher chances of locoregional recurrence and shorter survival in patients with HNSCC. [56] High expression level of miR-205 can be used to detect HNSCC lymph node metastasis. [99] A previous study identified lower expression level of miR-451 in HNSCC tumors as a strong predictor of recurrence of the tumor. [100] Moreover, in some cancers such as lung cancer, existence of cancer-specific miR-NAs can diagnose the cancer and classify the tumor subtype. [101]

Additionally, miRNAs may have an impact on the analysis of surgical margins for residual cancer extension. [102] High endogenous expression of mir-205 in normal and HNSCC cells and its low expression in lymph node tissue makes it a worthy biomarker for detecting a small number of tumor cells in the metastatic site. [99] Taken together, these findings show that altered expression of miRNAs could be a promising marker for HNSCC prognosis.

Discussion

Due to a gradual increase in the number of cancer patients, careful evaluation of patients has a great impact on decreasing the cancer-related morbidity and mortality rate. Among the cancer groups, HNSCCs consist of highly heterogeneous lesions. [103]

It seems that in the metastatic phenotype of miR

NAs, a global reduction of miRNA abundance plays a causal role; [11] however; several miRNAs are upregulated in specific tumors. Moreover, miRNAs act as either oncogenes or tumor suppressors which make them prospective targets for therapeutic development against cancers. [104]

There are many challenges in the application of miRNA-based therapies. [32] For instance, miR-1 suppresses metastasis in HNSCC by targeting TAGLN2; [53] however, targeting MET, a proto-oncogene, is the underlying mechanism for anti-tumorigenic property of miR-1 in lung cancer.

Similarly, miR-1 down-regulation was observed in other types of cancer such as lung cancer. [105] A previous study indicated that miR-29 was downregulated in breast cancer; miR-29 has an essential role in inhibiting growth of breast cancer cells via downregulation of B-Myb. [106] But in HNSCC tissue, the target genes of mir-29 are LAMC2 and ITGA6. [58] The miR-34 family may inhibit invasion and metastasis in different cancer types. For example, miR-34a and b are suggested as tumor suppressors in several cancers such as gastric cancer, [107] breast cancer, [108] and hepatocellular carcinoma mainly by targeting p53. [109] mir34a/c may function as a metastasis suppressor in breast cancer through targeting Fos-related antigen 1 (FOSL1). [110] Nonetheless, the target gene of miR-34 in HNSCC is not clear. [63] miR-133a acts as a tumor suppressor in esophageal squamous cell carcinoma by targeting Fascin Actin-Bundling Protein 1 (FSCN1), an actin bundling protein which promotes the degradation of ECM, and matrix metalloproteinase 14 (MMP14). [111] But the target gene of miR-133a in HNSCC is MSN. [67] miR-99 functions as a tumor suppressor in some cancers via different targets. For example in HNSCC, it acts via IGFIR and mTOR, though in cervical cancer the target gene is Tribbles (TRIB2) which controls the specificity of the activation of mitogenactivated protein kinases (MAPK). [112] miR-138 also targets different genes in human colorectal cancer (Twist 2) compared with HNSCC (GNAI2, RhoC, and ROCK2). [113] miR-10b was the first link between metastasis and miRNAs indicated in breast cancer. Twist which induces EMT is a target of miR-10b. [114] Moreover, miR-10b targets E-cadherin and modulates metastasis in breast cancer; [115] however; its target

gene in HNSCC is still unknown. The literature shows that miR-21 is one of the most studied miRNAs. [116] Although miR-21 functions as an oncogene via programmed cell death 4(PDCD4) in HNSCC, [81] and esophageal carcinoma, [117] it stimulates cell invasion and promotes metastasis in breast cancer by targeting both PDCD4, and tropomyosin 1 (TPM1), a tumor suppressor gene. [118-119] On the other hand, miR-21 controls esophageal squamous cell carcinoma invasion by targeting the tissue inhibitor of matrix metalloproteinase-3 (TIMP3) molecules which control ECM. [120] miRNAs expression pattern and function are cancertype-specific. For example, up-regulation of miR-31 has been reported in colorectal cancer; [121] however, reduced level of miR-31 was observed in breast cancer. [122] While it inhibits metastasis in breast cancer, [123] it induces invasion and metastasis in colon cancer via TGF-B. [124] In addition, miR-181a and mir-181b suppress glioma tumor growth, [125] but contributes to tumor progression in thyroid papillary carcinoma. [126] miR-211 acts as a tumor suppressor in melanoma, [127] whereas; it functions as an oncomir in colon cancer. [128] Up-regulation of miR-222 in gastric carcinoma is associated with lymph node metastasis, vascular invasion and TNM stage, [129] but miR-222 regulates OSCC as a tumor suppressor and inhibits tumor invasion and metastasis. [12] MiR-223 acts as an oncogene in gastric cancer, and enhances cancer invasion and metastasis by targeting tumor suppressor genes F-box and WD40 domain protein 7 (FBXW7) and erythrocyte membrane protein band 4.1-like 3 (EPB41L3). But, the target gene of miR-223 in HNSCC is not clear. [63] miR-363 is a tumor suppressor in T cell lymphoma, [130] however; it acts as an oncogene in breast cancer. [131]

Conclusion

In conclusion, despite advances in cancer treatment, the mortality rate of HNSCC is still high. The potential application of micro-RNAs (miRNAs) in cancer therapy has been demonstrated in many studies; miRNAs function as either tumor suppressor or oncogene. The recognition of metastamir and their targets may lead to better understanding of HNSCC oncogenesis, and consequently the development of new therapeutic strategies for HNSCC which is a necessity in cancer treatment. Development of therapeutic agents based on miRNAs is a promising target.

Acknowledgement

This paper was supported by Hamadan University of Medical Sciences.

Conflict of Interest

The authors of this manuscript certify that they have no conflict of interest.

References

- Parkin DM, Bray F, Ferlay J, Pisani P. Global cancer statistics, 2002. CA Cancer J Clin. 2005; 55: 74-108.
- [2] Argiris A, Eng C. Epidemiology, staging, and screening of head and neck cancer. Cancer Treat Res. 2003; 114: 15-60.
- [3] Liu X, Chen Z, Yu J, Xia J, Zhou X. MicroRNA profiling and head and neck cancer. Comp Funct Genomics. 2009: 837514.
- [4] Chen D, Cabay RJ, Jin Y, Wang A, Lu Y, Shah-Khan M, Zhou X. Microrna deregulations in head and neck squamous cell carcinomas. J Oral Maxillofac Res. 2013; 4: e2.
- [5] Irani S. Metastasis to head and neck area: a 16-year retrospective study. Am J Otolaryngol. 2011; 32: 24-27.
- [6] Ellenrieder V, Adler G, Gress TM. Invasion and metastasis in pancreatic cancer. Ann Oncol. 1999; 10 Suppl 4: 46-50.
- [7] Takes RP. Staging of the neck in patients with head and neck squamous cell cancer: imaging techniques and biomarkers. Oral Oncol. 2004; 40: 656-667.
- [8] Kowalski LP, Bagietto R, Lara JR, Santos RL, Tagawa EK, Santos IR. Factors influencing contralateral lymph node metastasis from oral carcinoma. Head Neck. 1999; 21: 104-110.
- [9] Kowalski LP, Carvalho AL, Martins Priante AV, Magrin J. Predictive factors for distant metastasis from oral and oropharyngeal squamous cell carcinoma. Oral Oncol. 2005; 41: 534-541.
- [10] Nagadia R, Pandit P, Coman WB, Cooper-White J, Punyadeera C. miRNAs in head and neck cancer revisited. Cell Oncol (Dordr). 2013; 36: 1-7.
- [11] Lu J, Getz G, Miska EA, Alvarez-Saavedra E, Lamb J, Peck D, et al. MicroRNA expression profiles classify human cancers. Nature. 2005; 435: 834-838.

- [12] Liu X, Yu J, Jiang L, Wang A, Shi F, Ye H, et al. MicroRNA-222 regulates cell invasion by targeting matrix metalloproteinase 1 (MMP1) and manganese superoxide dismutase 2 (SOD2) in tongue squamous cell carcinoma cell lines. Cancer Genomics Proteomics. 2009; 6: 131-139.
- [13] Wójcicka A, Kolanowska M, Jażdżewski K. Mechanisms in endocrinology: microrna in diagnostics and therapy of thyroid cancer. Eur J Endocrinol. 2016; 174: R89-R98.
- [14] McGuire A, Brown JA, Kerin MJ. Metastatic breast cancer: the potential of miRNA for diagnosis and treatment monitoring. Cancer Metastasis Rev. 2015; 34: 145-155.
- [15] Cai X, Hagedorn CH, Cullen BR. Human microRNAs are processed from capped, polyadenylated transcripts that can also function as mRNAs. RNA. 2004; 10: 1957-1966.
- [16] Daley TD, Minett CP, Driman DK, Darling MR. Oral metastatic hepatocellular carcinoma: a changing demographic in Europe and North America. Immunohistochemical advances in the microscopic diagnosis. Oral Oncol. 2011; 47: 62-67.
- [17] Bushati N, Cohen SM. microRNA functions. Annu Rev Cell Dev Biol. 2007; 23: 175-205.
- [18] Bohnsack MT, Czaplinski K, Gorlich D. Exportin 5 is a RanGTP-dependent dsRNA-binding protein that mediates nuclear export of pre-miRNAs. RNA. 2004; 10: 185-191.
- [19] Chendrimada TP, Gregory RI, Kumaraswamy E, Norman J, Cooch N, Nishikura K, et al. TRBP recruits the Dicer complex to Ago2 for microRNA processing and gene silencing. Nature. 2005; 436(7051): 740-744.
- [20] Lewis BP, Shih IH, Jones-Rhoades MW, Bartel DP, Burge CB. Prediction of mammalian microRNA targets. Cell. 2003; 115: 787-798.
- [21] Hutvágner G, Zamore PD. A microRNA in a multipleturnover RNAi enzyme complex. Science. 2002; 297(5589): 2056-2060.
- [22] Olsen PH, Ambros V. The lin-4 regulatory RNA controls developmental timing in Caenorhabditis elegans by blocking LIN-14 protein synthesis after the initiation of translation. Dev Biol. 1999; 216: 671-680.
- [23] Bartels CL, Tsongalis GJ. MicroRNAs: novel biomarkers for human cancer. Clin Chem. 2009; 55: 623-631.
- [24] Ramdas L, Giri U, Ashorn CL, Coombes KR, El-Naggar A, Ang KK, et al. miRNA expression profiles in head and neck squamous cell carcinoma and adjacent normal

tissue. Head Neck. 2009; 31: 642-654.

- [25] Volinia S, Calin GA, Liu CG, Ambs S, Cimmino A, Petrocca F, et al. A microRNA expression signature of human solid tumors defines cancer gene targets. Proc Natl Acad Sci U S A. 2006; 103: 2257-2261.
- [26] Yamamoto E, Miyakawa A, Kohama G. Mode of invasion and lymph node metastasis in squamous cell carcinoma of the oral cavity. Head Neck Surg. 1984; 6: 938-947.
- [27] Kalluri R, Weinberg RA. The basics of epithelialmesenchymal transition. J Clin Invest. 2009; 119: 1420-1428.
- [28] Yang J, Weinberg RA. Epithelial-mesenchymal transition: at the crossroads of development and tumor metastasis. Dev Cell. 2008; 14: 818-829.
- [29] Zhang J, Ma L. MicroRNA control of epithelialmesenchymal transition and metastasis. Cancer Metastasis Rev. 2012; 31: 653-662.
- [30] Wang L, Song G, Tan W, Qi M, Zhang L, Chan J, et al. MiR-573 inhibits prostate cancer metastasis by regulating epithelial-mesenchymal transition. Oncotarget. 2015; 6: 35978-35990.
- [31] Qu J, Li M, An J, Zhao B, Zhong W, Gu Q, et al. MicroRNA-33b inhibits lung adenocarcinoma cell growth, invasion, and epithelial-mesenchymal transition by suppressing Wnt/β-catenin/ZEB1 signaling. Int J Oncol. 2015; 47: 2141-2152.
- [32] Zhao X, Dou W, He L, Liang S, Tie J, Liu C, et al. MicroRNA-7 functions as an anti-metastatic microRNA in gastric cancer by targeting insulin-like growth factor-1 receptor. Oncogene. 2013; 32: 1363-1372.
- [33] Long H, Wang Z, Chen J, Xiang T, Li Q, Diao X, et al. microRNA-214 promotes epithelial-mesenchymal transition and metastasis in lung adenocarcinoma by targeting the suppressor-of-fused protein (Sufu). Oncotarget. 2015; 6: 38705-38718.
- [34] Yanaka Y, Muramatsu T, Uetake H, Kozaki K, Inazawa J. miR-544a induces epithelial-mesenchymal transition through the activation of WNT signaling pathway in gastric cancer. Carcinogenesis. 2015; 36: 1363-1371.
- [35] Irani S, Salajegheh A, Smith RA, Lam AK. A review of the profile of endothelin axis in cancer and its management. Crit Rev Oncol Hematol. 2014; 89: 314-321.
- [36] Irani S, Salajegheh A, Gopalan V, Smith RA, Lam AK. Expression profile of endothelin 1 and its receptor endothelin receptor A in papillary thyroid carcinoma and their

correlations with clinicopathologic characteristics. Ann Diagn Pathol. 2014; 18: 43-48.

- [37] Zhu X, Li H, Long L, Hui L, Chen H, Wang X, et al. miR-126 enhances the sensitivity of non-small cell lung cancer cells to anticancer agents by targeting vascular endothelial growth factor A. Acta Biochim Biophys Sin (Shanghai). 2012; 44: 519-526.
- [38] Dews M, Homayouni A, Yu D, Murphy D, Sevignani C, Wentzel E, et al. Augmentation of tumor angiogenesis by a Myc-activated microRNA cluster. Nat Genet. 2006; 38: 1060-1065.
- [39] Bonauer A, Carmona G, Iwasaki M, Mione M, Koyanagi M, Fischer A, et al. MicroRNA-92a controls angiogenesis and functional recovery of ischemic tissues in mice. Science. 2009; 324: 1710-1713.
- [40] Rather MI, Nagashri MN, Swamy SS, Gopinath KS, Kumar A. Oncogenic microRNA-155 down-regulates tumor suppressor CDC73 and promotes oral squamous cell carcinoma cell proliferation: implications for cancer therapeutics. J Biol Chem. 2013; 288: 608-618.
- [41] Henson BJ, Bhattacharjee S, O'Dee DM, Feingold E, Gollin SM. Decreased expression of miR-125b and miR-100 in oral cancer cells contributes to malignancy. Genes Chromosomes Cancer. 2009; 48: 569-582.
- [42] Hauser B, Zhao Y, Pang X, Ling Z, Myers E, Wang P, et al. Functions of MiRNA-128 on the regulation of head and neck squamous cell carcinoma growth and apoptosis. PLoS One. 2015; 10(3): e0116321.
- [43] Coutinho-Camillo CM, Lourenço SV, de Araújo Lima L, Kowalski LP, Soares FA. Expression of apoptosisregulating miRNAs and target mRNAs in oral squamous cell carcinoma. Cancer Genet. 2015; 208: 382-389.
- [44] Park SY, Lee JH, Ha M, Nam JW, Kim VN. miR-29 miRNAs activate p53 by targeting p85 alpha and CDC42. Nat Struct Mol Biol. 2009; 16: 23-29.
- [45] Lynam-Lennon N, Maher SG, Reynolds JV. The roles of microRNA in cancer and apoptosis. Biol Rev Camb Philos Soc. 2009; 84: 55-71.
- [46] Wang J, Huang H, Wang C, Liu X, Hu F, Liu M. MicroRNA-375 sensitizes tumour necrosis factor-alpha (TNF-α)-induced apoptosis in head and neck squamous cell carcinoma in vitro. Int J Oral Maxillofac Surg. 2013; 42: 949-955.
- [47] Cervigne NK, Reis PP, Machado J, Sadikovic B, Bradley G, Galloni NN, et al. Identification of a microRNA signature associated with progression of leukoplakia to oral

carcinoma. Hum Mol Genet. 2009; 18: 4818-4829.

- [48] Budhu A, Jia HL, Forgues M, Liu CG, Goldstein D, Lam A, et al. Identification of metastasis-related microRNAs in hepatocellular carcinoma. Hepatology. 2008; 47: 897-907.
- [49] Giordano S, Columbano A. MicroRNAs: new tools for diagnosis, prognosis, and therapy in hepatocellular carcinoma? Hepatology. 2013; 57: 840-847.
- [50] Zhang B, Pan X, Cobb GP, Anderson TA. microRNAs as oncogenes and tumor suppressors. Dev Biol. 2007; 302: 1-12.
- [51] Calin GA, Dumitru CD, Shimizu M, Bichi R, Zupo S, Noch E, et al. Frequent deletions and down-regulation of micro- RNA genes miR15 and miR16 at 13q14 in chronic lymphocytic leukemia. Proc Natl Acad Sci U S A. 2002; 99: 15524-15529.
- [52] Hurst DR, Edmonds MD, Welch DR. Metastamir: the field of metastasis-regulatory microRNA is spreading. Cancer Res. 2009; 69: 7495-7498.
- [53] Nohata N, Sone Y, Hanazawa T, Fuse M, Kikkawa N, Yoshino H, et al. miR-1 as a tumor suppressive microRNA targeting TAGLN2 in head and neck squamous cell carcinoma. Oncotarget. 2011; 2: 29-42.
- [54] Johnson SM, Grosshans H, Shingara J, Byrom M, Jarvis R, Cheng A, et al. RAS is regulated by the let-7 microRNA family. Cell. 2005; 120: 635-647.
- [55] Chang CJ, Hsu CC, Chang CH, Tsai LL, Chang YC, Lu SW, et al. Let-7d functions as novel regulator of epithelial-mesenchymal transition and chemoresistant property in oral cancer. Oncol Rep. 2011; 26: 1003-1010.
- [56] Childs G, Fazzari M, Kung G, Kawachi N, Brandwein-Gensler M, McLemore M, et al. Low-level expression of microRNAs let-7d and miR-205 are prognostic markers of head and neck squamous cell carcinoma. Am J Pathol. 2009; 174: 736-745.
- [57] Chang CC, Yang YJ, Li YJ, Chen ST, Lin BR, Wu TS, et al. MicroRNA-17/20a functions to inhibit cell migration and can be used a prognostic marker in oral squamous cell carcinoma. Oral Oncol. 2013; 49: 923-931.
- [58] Kinoshita T, Nohata N, Hanazawa T, Kikkawa N, Yamamoto N, Yoshino H, et al. Tumour-suppressive microRNA-29s inhibit cancer cell migration and invasion by targeting laminin-integrin signalling in head and neck squamous cell carcinoma. Br J Cancer. 2013; 109: 2636-2645.
- [59] Heldin CH, Vanlandewijck M, Moustakas A. Regulation

of EMT by TGF β in cancer. FEBS Lett. 2012; 586: 1959-1970.

- [60] Roderburg C, Urban GW, Bettermann K, Vucur M, Zimmermann H, Schmidt S, et al. Micro-RNA profiling reveals a role for miR-29 in human and murine liver fibrosis. Hepatology. 2011; 53: 209-218.
- [61] Siemens H, Jackstadt R, Hünten S, Kaller M, Menssen A, Götz U, et al. miR-34 and SNAIL form a double-negative feedback loop to regulate epithelial-mesenchymal transitions. Cell Cycle. 2011; 10: 4256-4271.
- [62] Wong TS, Liu XB, Wong BY, Ng RW, Yuen AP, Wei WI. Mature miR-184 as Potential Oncogenic microRNA of Squamous Cell Carcinoma of Tongue. Clin Cancer Res. 2008; 14: 2588-2592.
- [63] Chen D, Cabay RJ, Jin Y, Wang A, Lu Y, Shah-Khan M, Zhou X. MicroRNA Deregulations in Head and Neck Squamous Cell Carcinomas. J Oral Maxillofac Res. 2013; 4: e2.
- [64] Chen Z, Jin Y, Yu D, Wang A, Mahjabeen I, Wang C, et al. Down-regulation of the microRNA-99 family members in head and neck squamous cell carcinoma. Oral Oncol. 2012; 48: 686-691.
- [65] Xu N, Zhang L, Meisgen F, Harada M, Heilborn J, Homey B, et al. MicroRNA-125b down-regulates matrix metallopeptidase 13 and inhibits cutaneous squamous cell carcinoma cell proliferation, migration, and invasion. J Biol Chem. 2012; 287: 29899-29908.
- [66] Sasahira T, Kurihara M, Bhawal UK, Ueda N, Shimomoto T, Yamamoto K, et al. Downregulation of miR-126 induces angiogenesis and lymphangiogenesis by activation of VEGF-A in oral cancer. Br J Cancer. 2012; 107: 700-706.
- [67] Kinoshita T, Nohata N, Fuse M, Hanazawa T, Kikkawa N, Fujimura L, et al. Tumor suppressive microRNA-133a regulates novel targets: moesin contributes to cancer cell proliferation and invasion in head and neck squamous cell carcinoma. Biochem Biophys Res Commun. 2012; 418: 378-383.
- [68] Jiang L, Dai Y, Liu X, Wang C, Wang A, Chen Z, et al. Identification and experimental validation of G protein alpha inhibiting activity polypeptide 2 (GNAI2) as a microRNA-138 target in tongue squamous cell carcinoma. Hum Genet. 2011; 129: 189-197.
- [69] Liu X, Jiang L, Wang A, Yu J, Shi F, Zhou X. MicroRN-A-138 suppresses invasion and promotes apoptosis in head and neck squamous cell carcinoma cell lines. Canc-

er Lett. 2009; 286: 217-222.

- [70] Liu X, Wang C, Chen Z, Jin Y, Wang Y, Kolokythas A, et al. MicroRNA-138 suppresses epithelial-mesenchymal transition in squamous cell carcinoma cell lines. Biochem J. 2011; 440: 23-31.
- [71] Jiang L, Liu X, Kolokythas A, Yu J, Wang A, Heidbreder CE, et al. Downregulation of the Rho GTPase signaling pathway is involved in the microRNA-138-mediated inhibition of cell migration and invasion in tongue squamous cell carcinoma. Int J Cancer. 2010; 127: 505-512.
- [72] Xu Q, Sun Q, Zhang J, Yu J, Chen W, Zhang Z. Downregulation of miR-153 contributes to epithelialmesenchymal transition and tumor metastasis in human epithelial cancer. Carcinogenesis. 2013; 34: 539-549.
- [73] Tamagawa S, Beder LB, Hotomi M, Gunduz M, Yata K, Grenman R, et al. Role of miR-200c/miR-141 in the regulation of epithelial-mesenchymal transition and migration in head and neck squamous cell carcinoma. Int J Mol Med. 2014; 33: 879-886.
- [74] Lo WL, Yu CC, Chiou GY, Chen YW, Huang PI, Chien CS, et al. MicroRNA-200c attenuates tumour growth and metastasis of presumptive head and neck squamous cell carcinoma stem cells. J Pathol. 2011; 223: 482-495.
- [75] Liu X, Yu J, Jiang L, Wang A, Shi F, Ye H, et al. MicroRNA-222 regulates cell invasion by targeting matrix metalloproteinase 1 (MMP1) and manganese superoxide dismutase 2 (SOD2) in tongue squamous cell carcinoma cell lines. Cancer Genomics Proteomics. 2009; 6: 131-139.
- [76] Sun Q, Zhang J, Cao W, Wang X, Xu Q, Yan M, et al. Dysregulated miR-363 affects head and neck cancer invasion and metastasis by targeting podoplanin. Int J Biochem Cell Biol. 2013; 45: 513-520.
- [77] Harris T, Jimenez L, Kawachi N, Fan JB, Chen J, Belbin T, et al. Low-level expression of miR-375 correlates with poor outcome and metastasis while altering the invasive properties of head and neck squamous cell carcinomas. Am J Pathol. 2012; 180: 917-928.
- [78] Siow MY, Ng LP, Vincent-Chong VK, Jamaludin M, Abraham MT, Abdul Rahman ZA, et al. Dysregulation of miR-31 and miR-375 expression is associated with clinical outcomes in oral carcinoma. Oral Dis. 2014; 20: 345-351.
- [79] Severino P, Brüggemann H, Andreghetto FM, Camps C, Klingbeil Mde F, de Pereira WO, et al. MicroRNA expression profile in head and neck cancer: HOX-cluster

Irani S.

embedded microRNA-196a and microRNA-10b dysregulation implicated in cell proliferation. BMC Cancer. 2013; 13: 533.

- [80] Lu YC, Chen YJ, Wang HM, Tsai CY, Chen WH, Huang YC, et al. Oncogenic function and early detection potential of miRNA-10b in oral cancer as identified by microRNA profiling. Cancer Prev Res (Phila). 2012; 5: 665-674.
- [81] Reis PP, Tomenson M, Cervigne NK, Machado J, Jurisica I, Pintilie M, et al. Programmed cell death 4 loss increases tumor cell invasion and is regulated by miR-21 in oral squamous cell carcinoma. Mol Cancer. 2010; 9: 238.
- [82] Li J, Huang H, Sun L, Yang M, Pan C, Chen W, et al. MiR-21 indicates poor prognosis in tongue squamous cell carcinomas as an apoptosis inhibitor. Clin Cancer Res. 2009; 15: 3998-4008.
- [83] Hedbäck N, Jensen DH, Specht L, Fiehn AM, Therkildsen MH, Friis-Hansen L, et al. MiR-21 expression in the tumor stroma of oral squamous cell carcinoma: an independent biomarker of disease free survival. PLoS One. 2014; 9: e95193.
- [84] Liu CJ, Tsai MM, Hung PS, Kao SY, Liu TY, Wu KJ, et al. miR-31 ablates expression of the HIF regulatory factor FIH to activate the HIF pathway in head and neck carcinoma. Cancer Res. 2010; 70: 1635-1644.
- [85] Li G, Ren S, Su Z, Liu C, Deng T, Huang D, et al. Increased expression of miR-93 is associated with poor prognosis in head and neck squamous cell carcinoma. Tumour Biol. 2015; 36: 3949-3956.
- [86] Smith AL, Iwanaga R, Drasin DJ, Micalizzi DS, Vartuli RL, Tan AC, et al. The miR-106b-25 cluster targets Smad7, activates TGF-β signaling, and induces EMT and tumor initiating cell characteristics downstream of Six1 in human breast cancer. Oncogene. 2012; 31: 5162-5171.
- [87] Fang L, Deng Z, Shatseva T, Yang J, Peng C, Du WW, et al. MicroRNA miR-93 promotes tumor growth and angiogenesis by targeting integrin-β8. Oncogene. 2011; 30: 806-821.
- [88] Liu CJ, Shen WG, Peng SY, Cheng HW, Kao SY, Lin SC, et al. miR-134 induces oncogenicity and metastasis in head and neck carcinoma through targeting WWOX gene. Int J Cancer. 2014; 134: 811-821.
- [89] Ni YH, Huang XF, Wang ZY, Han W, Deng RZ, Mou YB, et al. Upregulation of a potential prognostic biomarker, miR-155, enhances cell proliferation in patients with oral squamous cell carcinoma. Oral Surg Oral Med

Oral Pathol Oral Radiol. 2014; 117: 227-233.

- [90] Baba O, Hasegawa S, Nagai H, Uchida F, Yamatoji M, Kanno NI, et al. MicroRNA-155-5p is associated with oral squamous cell carcinoma metastasis and poor prognosis. J Oral Pathol Med. 2015. doi: 10.1111/jop.12351. [Epub ahead of print]
- [91] Lajer CB, Nielsen FC, Friis-Hansen L, Norrild B, Borup R, Garnæs E, et al. Different miRNA signatures of oral and pharyngeal squamous cell carcinomas: a prospective translational study. Br J Cancer. 2011; 104: 830-840.
- [92] Chu TH, Yang CC, Liu CJ, Lui MT, Lin SC, Chang KW. miR-211 promotes the progression of head and neck carcinomas by targeting TGFβRII. Cancer Lett. 2013; 337: 115-124.
- [93] Chang KW, Liu CJ, Chu TH, Cheng HW, Hung PS, Hu WY, et al. Association between high miR-211 microRNA expression and the poor prognosis of oral carcinoma. J Dent Res. 2008; 87: 1063-1068.
- [94] Park NJ, Zhou H, Elashoff D, Henson BS, Kastratovic DA, Abemayor E, et al. Salivary microRNA: discovery, characterization, and clinical utility for oral cancer detection. Clin Cancer Res. 2009; 15: 5473-5477.
- [95] Avissar M, Christensen BC, Kelsey KT, Marsit CJ. MicroRNA expression ratio is predictive of head and neck squamous cell carcinoma. Clin Cancer Res. 2009; 15: 2850-2855.
- [96] Bloomston M, Frankel WL, Petrocca F, Volinia S, Alder H, Hagan JP, et al. MicroRNA expression patterns to differentiate pancreatic adenocarcinoma from normal pancreas and chronic pancreatitis. JAMA. 2007; 297: 1901-1908.
- [97] Guo Y, Chen Z, Zhang L, Zhou F, Shi S, Feng X, et al. Distinctive microRNA profiles relating to patient survival in esophageal squamous cell carcinoma. Cancer Res. 2008; 68: 26-33.
- [98] Barker EV, Cervigne NK, Reis PP, Goswami RS, Xu W, Weinreb I, et al. microRNA evaluation of unknown primary lesions in the head and neck. Mol Cancer. 2009; 8: 127.
- [99] Fletcher AM, Heaford AC, Trask DK. Detection of metastatic head and neck squamous cell carcinoma using the relative expression of tissue-specific mir-205. Transl Oncol. 2008; 1: 202-208.
- [100] Hui AB, Lenarduzzi M, Krushel T, Waldron L, Pintilie M, Shi W, et al. Comprehensive MicroRNA profiling for head and neck squamous cell carcinomas. Clin Canc-

er Res. 2010; 16: 1129-1139.

- [101] Landi MT, Zhao Y, Rotunno M, Koshiol J, Liu H, Bergen AW, et al. MicroRNA expression differentiates histology and predicts survival of lung cancer. Clin Cancer Res. 2010; 16: 430-441.
- [102] Santhi WS, Prathibha R, Charles S, Anurup KG, Reshmi G, Ramachandran S, et al. Oncogenic microRNAs as biomarkers of oral tumorigenesis and minimal residual disease. Oral Oncol. 2013; 49: 567-575.
- [103] Siegel R, Naishadham D, Jemal A. Cancer statistics, 2012. CA Cancer J Clin. 2012; 62: 10-29.
- [104] Chen LH, Tsai KL, Chen YW, Yu CC, Chang KW, Chiou SH, et al. MicroRNA as a Novel Modulator in Head and Neck Squamous Carcinoma. J Oncol. 2010; 2010: 135632.
- [105] Nasser MW, Datta J, Nuovo G, Kutay H, Motiwala T, Majumder S, et al. Down-regulation of micro-RNA-1 (miR-1) in lung cancer. Suppression of tumorigenic property of lung cancer cells and their sensitization to doxorubicin-induced apoptosis by miR-1. J Biol Chem. 2008; 283: 33394-33405.
- [106] Wu Z, Huang X, Huang X, Zou Q, Guo Y. The inhibitory role of Mir-29 in growth of breast cancer cells. J Exp Clin Cancer Res. 2013; 32: 98.
- [107] Ji Q, Hao X, Meng Y, Zhang M, Desano J, Fan D, et al. Restoration of tumor suppressor miR-34 inhibits human p53-mutant gastric cancer tumorspheres. BMC Cancer. 2008; 8: 266.
- [108] Lee YM, Lee JY, Ho CC, Hong QS, Yu SL, Tzeng CR, et al. miRNA-34b as a tumor suppressor in estrogendependent growth of breast cancer cells. Breast Cancer Res. 2011; 13: R116.
- [109] Li N, Fu H, Tie Y, Hu Z, Kong W, Wu Y, et al. miR-34a inhibits migration and invasion by down-regulation of c-Met expression in human hepatocellular carcinoma cells. Cancer Lett. 2009; 275: 44-53.
- [110] Yang S, Li Y, Gao J, Zhang T, Li S, Luo A, et al. MicroRNA-34 suppresses breast cancer invasion and metastasis by directly targeting Fra-1. Oncogene. 2013; 32: 4294-4303.
- [111] Akanuma N, Hoshino I, Akutsu Y, Murakami K, Isozaki Y, Maruyama T, et al. MicroRNA-133a regulates the mRNAs of two invadopodia-related proteins, FSCN1 and MMP14, in esophageal cancer. Br J Cancer. 2014; 110: 189-198.
- [112] Xin JX, Yue Z, Zhang S, Jiang ZH, Wang PY, Li YJ, et

al. miR-99 inhibits cervical carcinoma cell proliferation by targeting TRIB2. Oncol Lett. 2013; 6: 1025-1030.

- [113] Long L, Huang G, Zhu H, Guo Y, Liu Y, Huo J. Downregulation of miR-138 promotes colorectal cancer metastasis via directly targeting TWIST2. J Transl Med. 2013; 11: 275.
- [114] Ma L, Teruya-Feldstein J, Weinberg RA. Tumour invasion and metastasis initiated by microRNA-10b in breast cancer. Nature. 2007; 449: 682-688.
- [115] Liu Y, Zhao J, Zhang PY, Zhang Y, Sun SY, Yu SY, et al. MicroRNA-10b targets E-cadherin and modulates breast cancer metastasis. Med Sci Monit. 2012; 18: BR299-BR308.
- [116] Bose D, Nahar S, Rai MK, Ray A, Chakraborty K, Maiti S. Selective inhibition of miR-21 by phage display screened peptide. Nucleic Acids Res. 2015; 43: 4342-4352.
- [117] Liu T, Liu Q, Zheng S, Gao X, Lu M, Yang C, et al. MicroRNA-21 promotes cell growth and migration by targeting programmed cell death 4 gene in Kazakh's esophageal squamous cell carcinoma. Dis Markers. 2014; 2014: 232837.
- [118] Zhu S, Si ML, Wu H, Mo YY. MicroRNA-21 targets the tumor suppressor gene tropomyosin 1 (TPM1). J Biol Chem. 2007; 282: 14328-14336.
- [119] Zhu S, Wu H, Wu F, Nie D, Sheng S, Mo YY. MicroRNA-21 targets tumor suppressor genes in invasion and metastasis. Cell Res. 2008; 18: 350-359.
- [120] Wang N, Zhang CQ, He JH, Duan XF, Wang YY, Ji X, et al. MiR-21 down-regulation suppresses cell growth, invasion and induces cell apoptosis by targeting FASL, TIMP3, and RECK genes in esophageal carcinoma. Dig Dis Sci. 2013; 58: 1863-1870.
- [121] Bandrés E, Cubedo E, Agirre X, Malumbres R, Zárate R, Ramirez N, et al. Identification by Real-time PCR of 13 mature microRNAs differentially expressed in colorectal cancer and non-tumoral tissues. Mol Cancer. 2006; 5: 29.
- [122] Vimalraj S, Miranda PJ, Ramyakrishna B, Selvamurugan N. Regulation of breast cancer and bone metastasis by microRNAs. Dis Markers. 2013; 35: 369-387.
- [123] Valastyan S, Reinhardt F, Benaich N, Calogrias D, Szász AM, Wang ZC, et al. A pleiotropically acting microRNA, miR-31, inhibits breast cancer metastasis. Cell. 2009; 137: 1032-1046.
- [124] Cottonham CL, Kaneko S, Xu L. miR-21 and miR-31

converge on TIAM1 to regulate migration and invasion of colon carcinoma cells. J Biol Chem. 2010; 285: 35293-35302.

- [125] Shi L, Cheng Z, Zhang J, Li R, Zhao P, Fu Z, et al. hsamir-181a and hsa-mir-181b function as tumor suppressors in human glioma cells. Brain Res. 2008; 1236: 185-193.
- [126] Pallante P, Visone R, Ferracin M, Ferraro A, Berlingieri MT, Troncone G, et al. MicroRNA deregulation in human thyroid papillary carcinomas. Endocr Relat Cancer. 2006; 13: 497-508.
- [127] Xu Y, Brenn T, Brown ER, Doherty V, Melton DW. Differential expression of microRNAs during melanoma progression: miR-200c, miR-205 and miR-211 are downregulated in melanoma and act as tumour suppressors. Br J Cancer. 2012; 106: 553-561.
- [128] Liu Q, Huang J, Zhou N, Zhang Z, Zhang A, Lu Z, et al.

LncRNA loc285194 is a p53-regulated tumor suppressor. Nucleic Acids Res. 2013; 41: 4976-4987.

- [129] Wang M, Zhao C, Shi H, Zhang B, Zhang L, Zhang X, et al. Deregulated microRNAs in gastric cancer tissuederived mesenchymal stem cells: novel biomarkers and a mechanism for gastric cancer. Br J Cancer. 2014; 110: 1199-1210.
- [130] Ng SB, Yan J, Huang G, Selvarajan V, Tay JL, Lin B, et al. Dysregulated microRNAs affect pathways and targets of biologic relevance in nasal-type natural killer/T-cell lymphoma. Blood. 2011; 118: 4919-4929.
- [131] Beltran AS, Russo A, Lara H, Fan C, Lizardi PM, Blancafort P. Suppression of breast tumor growth and metastasis by an engineered transcription factor. PLoS One. 2011; 6: e24595.